

**ASPERGILLUS FLAVUS INFECTION AND AFLATOXIN
CONTAMINATION IN PEANUTS AT VARIOUS STAGES OF THE
DELIVERY CHAINS IN CIANJUR REGENCY, WEST JAVA, INDONESIA**

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ABSTRACT

A survey to obtain information on pre- and postharvest handling of peanuts at farmer, collector, wholesaler and retailer levels, including *Aspergillus flavus* infection and aflatoxin B₁ contamination of peanuts collected in Cianjur regency, West Java, was conducted during the harvest period of the wet season of February 2004. The moisture contents and physical qualities of the peanuts were also determined.

Thirteen and 40 dry pod samples were collected randomly from 12 farmers and 23 collectors, respectively. Seven dry kernel samples were also collected from collectors. Five and 45 dry kernel samples were collected randomly from 2 wholesalers and 45 retailers in traditional markets, respectively. Thus, a total of 110 dry peanut pod and kernel samples were collected.

The results of interviews with farmers, collectors, wholesalers and retailers, and also the moisture contents and physical qualities of the peanuts are described in this article.

The percentages of samples infected by *A. flavus* were highest at the wholesaler as well as at retailer levels (100%, respectively), followed by those sampled at the collectors (85.0 and 85.7%, respectively), and farmers (84.6%). The mean percentage of infected kernels in infected samples of peanuts collected from retailers was the highest (87.6%), followed by those collected from wholesalers (72.4%), collectors in the form of kernels (23.3%) and pods (17.7%), and farmers (15.2%).

The range of aflatoxin B₁ contents in peanut samples collected from farmers (dry pods), collectors (dry pods), wholesalers (dry pods and kernels) and retailers (dry kernels) were < 3.6 - 114.2, < 3.6 - 2999.5 and < 3.6 - 34.1, < 3.6 - 6065.9, and < 3.6 - 6073.0 ppb, respectively. The highest aflatoxin B₁ contents at the wholesaler and retailer levels were 6065.9 ppb (in one sample) and 6073.0 ppb (in one sample), respectively. The percentage of samples contaminated with more than 15 ppb of aflatoxin B₁ was the highest in peanuts collected from wholesalers (80.0% of samples), followed by retailers (75.6%), farmers (38.5%) and collectors (30.0 and 14.3%). In 1999 Codex Alimentarius Commission determined that the maximum total aflatoxin content in peanuts intended for further processing is 15 ppb, suggesting that an alarming proportion of peanuts throughout the Indonesian food chain are in excess of this maximum limit.

Key words : *Aspergillus flavus*/Aflatoxin/Peanuts/Cianjur regency

INTRODUCTION

Peanuts are next to maize and soybean as the most important secondary crop in Indonesia. Since Indonesia has a humid tropical climate, peanuts can easily be

infected by moulds (including *A. flavus*) under drought stress before harvest, during the drying phase in the field, or under poor storage conditions. Aflatoxin has been recognized as a human and domestic animal carcinogen, and is produced following infection of peanuts by certain strains of *A. flavus*.

Pitt and Hocking (1996) reported that 45% of 215 peanut samples collected from farm storage, middlemen and retailers in Bogor (West Java), Yogyakarta (Central Java), and their surroundings, contained more than 50 ppb of aflatoxin, 33% more than 300 ppb, and 22 % exceeded 1000 ppb.

The total annual cost of aflatoxins in peanuts in 1991 in Indonesia, Philippines and Thailand was estimated as about \$ A 158 million. Indonesia incurred 84% (= \$ A 132 million) of this cost (Lubulwa and Davis 1994).

According to Dharmaputra *et al.* (2003a), in general, aflatoxin BI contents of peanuts collected from farmer's fields/pewefoas/collectors and processed samples in the Pati regency of Central Java were low (less than 15 ppb). The highest aflatoxin BI contents were found in raw peanut kernels collected from retailers in traditional markets, ranging from 2-124 and < 4 - 342 ppb during the wet and dry seasons in 2002, respectively. The percentage of raw kernel samples contaminated with aflatoxin BI (exceeding 15 ppb) collected during the wet and dry seasons was 33 and 25%, respectively.

Another study was also carried out by Dharmaputra *et al.* (2005) on aflatoxin BI contents of peanuts collected from farmer's fields, collectors and retailers in the Wonogiri regency and specifically the city of Surakarta (Central Java) during the wet and dry seasons in 2003. The results also showed that the highest aflatoxin B₁ contents were found in raw peanut kernels collected from retailers in traditional markets, with the range of < 3.6 - 1859.3 and < 3.6 - 5511.5 ppb during the wet and dry seasons, respectively. The percentage of raw kernel samples contaminated with aflatoxin BI (exceeding 15 ppb) collected during the wet and dry seasons was 33 and 76 %, respectively.

The 23rd session of the Joint FAO/WHO Food Standards Programme held in Rome, Italy (28 June-3 July 1999) reported that Codex Alimentarius Commission has adopted a maximum level of total aflatoxins in peanuts intended for further processing at 15 ppb. On 9 September 2004 the National Agency for Drug and Food Control, Republic of Indonesia has determined that aflatoxin BI and total aflatoxin contents in processed peanut products should not be more than 20 and 35 ppb, respectively.

To minimize or to reduce aflatoxin contamination in peanuts, appropriate post-harvest handling methods in each level of peanut delivery chain (farmer, collector, wholesaler, and retailer) should be carried out.

The objective of this study was to obtain information on pre- and postharvest handling methods, *Aspergillus flavus* infection and aflatoxin BI contamination of peanuts collected from different points of the delivery chains in Cianjur regency of West Java. The moisture contents and physical quality of peanut kernels were also determined.

MATERIALS AND METHODS Time and location of surveys

Surveys were conducted during the harvest of the wet season (February 2004) at Cidaun, Naringgul and Sindangbarang districts located in Cianjur regency, and the city of Cianjur, West Java. Based on the information obtained from the Indonesian Government Regional Office for Food Crops of West Java Province, peanut production was high in Cianjur regency and it ranks second after Garut regency. Peanut production was high at Cidaun, Naringgul and Sindang Barang districts according to the information obtained from the Indonesian Government Regional Office for Food Crops of Cianjur regency.

The surveys comprised:

- interviews, using questionnaires, with farmers, collectors, wholesalers and retailers. The questionnaires consisted of questions relating to pre- and postharvest handling of peanuts.
- random sampling of various kinds of peanut products collected from farmers, collectors, wholesalers and retailers who were interviewed.

The moisture contents, physical quality of kernels, fungal (*A. flavus*) infection and aflatoxin B, contents from each sample were analyzed.

Sampling methods

Based on differences of storage duration, more than one peanut sample could be obtained from each farmer, collector and wholesaler. The kind of peanuts sampled included dry pods and kernels. Samples of dry pods (about 2 kg each) were divided three times manually and homogeneously to obtain working samples (about 250 g each) for analyzing moisture contents, physical quality of kernels, percentage of kernels infected by *A. flavus*, aflatoxin BI content, and a reserve sample. The dry peanut pods were then shelled manually.

Samples of dry peanut kernels (about 1 kg each) were also divided three times using a box divider to obtain working samples (about 125 g each) for analyzing moisture contents, physical quality of kernels, percentage of kernels infected by *A. flavus*, aflatoxin BI content, and a reserve sample.

Moisture content, physical quality of kernels, *A. flavus* and aflatoxin BI analyses

Moisture contents of kernels (based on a wet basis) were analyzed using a SINAR TM AP 6060 Moisture Analyzer. The moisture contents of some samples were confirmed using the oven method (BSI 1995). Two replicates were used from each sample.

Physical quality of kernels was assessed in intact, shriveled and damaged kernels. The damaged kernels included cracked, broken, discoloured, and damage caused by insects or fungi. The percentage of each category of kernels was determined by counting them and dividing the total number of kernels used for physical quality analysis.

The percentage of kernels infected by *A. flavus* was determined using a plating method (100 kernels per sample) on *Aspergillus flavus* and *parasiticus* agar (AFPA) (P'Metal. 1983).

Aflatoxin B_t was analyzed because it is the most dangerous toxin. Aflatoxin B, contents in the kernels were determined using the ELISA method (Lee and Kennedy 2002), with two replicates used for each sample.

RESULTS AND DISCUSSION Source, kind and number of samples

At the farmer level (12 farmers), 13 samples of dry peanut pods were collected, while at collector level (23 collectors), 40 samples of dry pods and 7 samples of dry kernels (1 kg/sample) were collected. At the wholesaler and retailer levels (2 wholesalers and 45 retailers), 5 and 45 samples of dry peanut kernels were collected, respectively. Thus, the total number of samples was 110. Details of the peanut delivery chain, location and number of peanut samples are presented in Table 1.

Table 1. Details of the peanut delivery chain, including location and number of peanut samples

Peanut delivery chain	Cianjur regency/ City of Cianjur	District	Traditional market	Number of sample
Farmer	Cianjur regency	Cidaun	-	13
Collector	Cianjur regency	Cidaun	-	16
		Naringgul	-	6
		Sindangbarang	-	25
Wholesaler	City of Cianjur	-	-	5
Retailer	Cianjur regency	-	Cipanas Market	3
			Sukancgara Market	3
			Bojong Meron Market	10
			Baru Market	2
			Muka Market	10
			Induk Market	17
			Total	110

Results of interviews with farmers, collectors, wholesalers and retailers concerning pre- and postharvest handling of peanuts

Interview with farmers

The results from farmer interviews (12 respondents) are presented in Table 2. All farmers (100%) planted a local variety of peanuts, with most of the seed sources from farmers (83% of respondents). Farmers harvested their peanuts at 90 to 100 days after planting. During planting, all of respondents used fertilizer, such as urea, TSP and K.C1. Weed control was conducted manually. Farmers sun-dried peanut pods for 3 - 4 days. Most of the farmers used woven polypropylene bags (67% of respondents) to dry peanuts. Based on their experiences, farmers could determine when their peanuts were fully dried for safe storage. Sixty seven percent of respondents stored peanuts in woven polypropylene bags for 1 - 7 days before selling to collectors. All respondents sold peanuts to collectors in the form of dry pods. Interestingly, all respondents were not aware of the aflatoxin problem in peanuts.

Table 2. Results of interviews with farmers on pre- and postharvest handling of peanuts in Cianjur regency, West Java

No	Subject	Percentage of respondents
1	Variety of peanuts:	
	a. Local	100
	b. Others	0
2	Seed source:	
	a. Belong to the farmers	83
	b. Farmers bought seeds	17
3	Using fertilizer:	
	a. Yes (Urea, TSP, KCl)	100
	b. No	0
4	Weed control:	
	a. Manually	100
	b. No control	0
5	Harvesting time of peanuts (days after planting):	
	a. 100	25
	b. 95	8
	c. 92	8
	d. 90	59
6	When the peanuts were harvested by farmers:	
	a. Method of harvesting:	
	• by pulling peanut plants manually	100
	• mechanically	0

Table 2. (continued)

No	Subject	Percentage of respondents
	b. Method of stripping the pods:	
	• manually	100
	• mechanical pod stripping	0
	c. Method of drying:	
	• sun-drying for:	
	- 3 days	33
	- 4 days	67
	• mechanical drying	0
7	Facility used for sun-drying the peanuts:	
	a. Bamboo mat	8
	b. Tarpaulin	25
	c. Woven polypropylene bag	67
8	Method used to obtain information that the peanuts were already dried enough:	
	a. Based on the experience	100
	b. Using moisture tester	0
9	Storing of peanuts before selling to collectors:	
	a. Yes	67
	b. No	33
10	The peanuts were stored in:	
	a. Woven polypropylene bags	100
	b. Others	0
11	Duration of storage:	
	a. 1 – 7 days	100
	b. > 7 days	0
12	After being stored the peanuts were sold/sent to:	
	a. Collectors in the form of wet pods	0
	b. Collectors in the form of dry pods	100
	c. Retailers in the form of wet pods/dry pods/kernels	0
13	Aware of aflatoxin in peanuts:	
	a. Yes	0
	b. No	100

Interview with collectors

The result of interview with collectors (23 respondents) are presented in Table 3. Based on their experience the collectors knew whether their peanuts were Fully dry. Before selling to wholesalers, peanuts were stored in woven polypropylene bags (91% respondents), or by spreading them on paved floor (9% of

respondents) for 1-30 days. Peanuts were sold to wholesalers in the form of dry pods (35% of respondents), or as dry pods and kernels to wholesalers and retailers (9% of respondents) in the city of Cianjur. Peanuts were also sold to wholesalers in the form of dry pods (43% of respondents), and to wholesalers and retailers in the form of kernels (4% of respondents) in the big cities of West, Central and East Java. All respondents shelled peanut pods using a diesel powered sheller. Most collectors (87% of respondents) sorted peanuts manually before selling them to wholesalers. All respondents were not aware of aflatoxin problem in peanuts.

Table 3. Results of interviews with collectors on postharvest handling of peanuts in Cianjur regency, West Java

No	Subject	Percentage of respondents
1	The peanuts were bought from:	
	a. Farmers in the form of dry pods	100
	b. Others	0
2	Measuring of moisture contents during drying:	
	a. Yes	0
	b. No	100
3	Method used to obtain information that the peanuts were already dried enough:	
	a. Based on the experience	100
	b. Using moisture tester	0
4	Storing of peanuts before being sent to retailers:	
	a. Yes	100
	b. No	0
5	The peanuts were stored in:	
	a. Polypropylene bags	91
	b. Spread on the paved floor	9
6	Duration of storage:	
	a. 1 - 7 days	39
	b. 1 - 7 days and > 7 - 30 days	13
	c. > 7 - 30 days	48
7	After being stored the peanuts were sold/sent to:	
	a. The city of Cianjur in the form of:	
	• dry pods to wholesalers	35
	• kernels to retailers	0
	• dry pods and kernels to wholesalers and retailers	9
	b. The big cities of West Java (Subang, Cirebon, Bandung, Jakarta), Central Java (Kudus, Pati, Jepara) and East Java (Tuban) in the form of:	
	• dry pods to wholesalers	43
	• kernels to retailers	4
	• dry pods and kernels to wholesalers and retailers	9

Table 3. (continued)

No	Subject	Percentage of respondents
8	Method of shelling the peanuts:	
	a. Manually	0
	b. Mechanical sheller made from wood or zinc	0
	c. Electric sheller	0
	d. Diesel powered sheller	100
9	Monitoring of physical quality of peanut kernels before being sold:	
	a. Yes (sorted manually)	87
	b. No	13
10	Aware of aflatoxin in peanuts:	
	a. Yes	0
	b. No	100

Interview with wholesalers

The results of interview with wholesalers (2 respondents) are presented in Table 4. At the wholesaler level peanuts were stored in woven polypropylene bags (50% of respondents), as well as in woven polypropylene and jute bags (50% of respondents). The stacks of bags containing peanuts were not placed on pallets. Peanuts samples were stored for 1 month (50% of respondents) and 1-2 months (50% of respondents). Fifty percents of the respondents sold only peanuts, while 50% of respondents sold peanuts and other commodities such as wheat flour and peanut oil. Peanuts were sold to smaller wholesalers in the cities of Bandung, Bogor and Jakarta.

Table 4. Results of interviews with **wholesalers** on postharvest handling of peanuts in the city of Cianjur, West Java

No	Subject	Percentage of respondents
1	The peanuts (in the form of pod or kernel) source:	
	a. Farmers	0
	b. Collector	100
2	The peanuts were stored in:	
	a. Woven polypropylene bag	50
	b. Jute bag	0
	c. Woven polypropylene and jute bags	50
3	The stack of bags containing peanuts were placed on pallets:	
	a. Yes	0
	b. No	100

Table 4. (continued)

No	Subject	Percentage of respondents
4	Storage duration of the samples	
	a. 1 month	50
	b. 2 months	0
	c. 1 - 2 months	50
5	Apart from peanuts, wholesaler sold other ingredients such as:	
	a. Wheat flour and peanut oils	50
	b. None	50
6	Peanut buyers were:	
	a. Smaller wholesalers in Bandung, Bogor and Jakarta	100
	b. Others	0

Interview with retailers

The results of interview with retailers (45 respondents) are presented in Table 5. Fifty three percent of respondents bought their peanuts from wholesalers, while 47% of respondent bought peanuts from farmers and collectors. Peanuts were stored in woven polypropylene bags (71% of respondents) and jute bags (29% of respondents) for 1 - 7 days. Containers used at the time of sampling were rectangular plastic basins (42% of respondents), winnowing trays (29% of respondents), wooden boxes (18% of respondents), round plastic basins (5% of respondents), woven polypropylene bags (4% of respondents) and jute bags (2% of respondents). Aside from peanuts, retailers also sold other general commodities. Peanut buyers included sellers of peanut sauce products for making *gado-gado*, *pecel* and *sate*, as well as house wives. Retailers were also not aware of the aflatoxin problem in peanuts.

Table 5. Results of interviews with **retailers** on postharvest handling of peanuts in Cianjur regency and in the city of Cianjur, West Java

No	Subject	Percentage of respondents
1	The peanuts source:	
	a. Farmers	22
	b. Collectors	25
	c. Wholesalers	53
2	The peanuts were stored in:	
	a. Woven polypropylene bag	71
	b. Jute bag	29

Table 5. Results of interviews with **retailers** on postharvest handling of peanuts in Cianjur regency and in the city of Cianjur, West Java

No	Subject	Percentage of respondents
3	Container used when peanut sampling was conducted:	
	a. Woven polypropylene bag	4
	b. Jute bag	2
	c. Round plastic basin	5
	d. Rectangular plastic basin	42
	e. Wooden box	18
	f. Winnowing tray	29
4	Storage duration of the samples:	
	a. 1 – 7 days	100
	b. > 7 days	0
5	Apart from peanuts, retailers sold:	
	a. Other general commodities	100
	b. Others	0
6	Peanut buyers were:	
	a. <i>gado-gado</i> , <i>pecel</i> and <i>sate</i> sellers and house wives	100
	b. Others	0

Moisture contents, physical quality of kernels, the incidence of *A. flavus* and aflatoxin contamination

Moisture contents

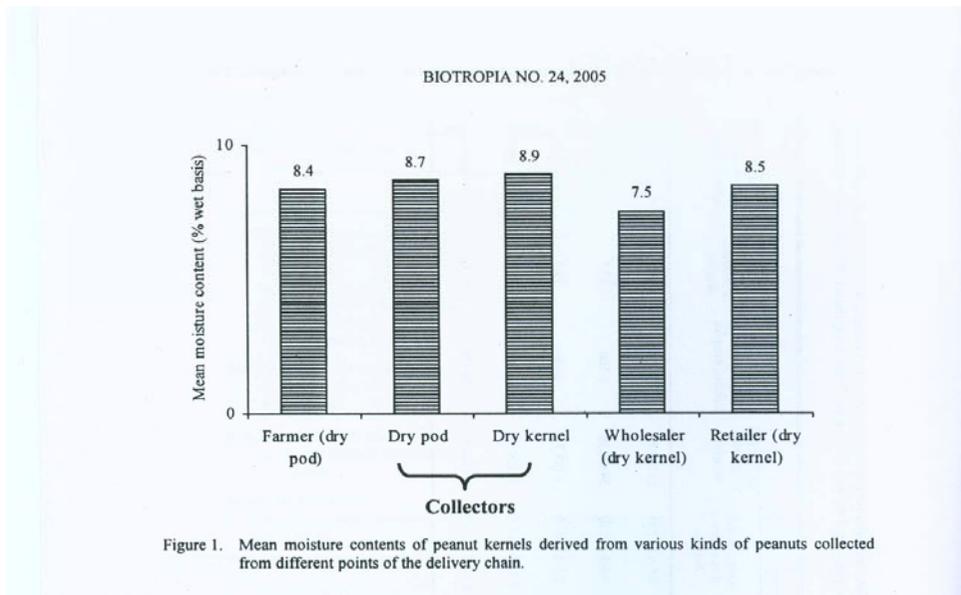
According to Diener and Davis (1969) moisture content is an important factor affecting the growth of *A. flavus* and aflatoxin production.

The range and mean moisture contents of kernels derived from various kinds of peanuts collected from farmers, collectors, wholesalers and retailers are presented in Table 6 and Figure 1. The moisture contents of kernels collected from farmers was relatively similar to those collected from collectors and retailers, while the moisture contents of kernels collected from wholesalers was the lowest. This was due to the storage duration of peanuts at farmer and collector levels which were relatively shorter than those at the wholesaler level (Tables 2, 3 and 4). The moisture contents of kernels were always in equilibrium with the relative humidity of storage room.

The range and mean moisture contents of kernels derived from various kinds of peanuts and collected from farmers, collectors, wholesalers and retailers were 5.9 - 9.9% and 8.4%; 7.8 -9.6% and 8.7%; 8.1 - 9.4% and 8.9%; 6.2 - 8.8% and 7.5%; 6.5 - 9.3% and 8.5%, respectively (Table 6). In general, the mean of moisture content of peanut kernels in each delivery chain were considered safe for storage. SNI (1995) determined that the safe moisture contents for storage of peanut pods and kernels were 9 and 8%, respectively.

Table 6. Moisture contents, physical quality of kernels, *Aspergillus flavus* infection, and aflatoxin B₁ content of peanut kernels derived from various kinds of peanuts collected from different points of the delivery chain in Cianjur regency and the city of Cianjur (West Java) during the wet season (February 2004)

Peanut delivery chain	Kind of peanuts	Number of samples	Range (mean) of moisture content (Based on % wet basis)	Physical quality of kernels					Number (%) of infected samples	Range of % infection in infected samples	Mean % of infected kernels in infected samples	Range of aflatoxin B ₁ content (ppb)
				Range (mean) of % intact kernels	Range (mean) of % shriveled kernels	Range (mean) of % damaged kernels	Range (mean) of % of intact kernels	Range (mean) of % damaged kernels				
Farmer	Dry pod	13	5.9 - 9.9 (8.4)	59.6-83.9 (74.8)	14.2-36.6 (22.9)	0.6-4.9 (2.3)	11	(84.6)	1-56	15.2	< 3.6 - 114.2	
Collector	Dry pod	40	7.8 - 9.6 (8.7)	42.2-83.0 (70.0)	13.7-56.4 (28.0)	0.6-5.6 (2.2)	34	(85.0)	1-100	17.7	< 3.6-2999.5	
	Raw kernel	7	8.1 - 9.4 (8.9)	56.2-77.3 (69.0)	19.7-41.7 (27.9)	2.1-4.5 (3.1)	6	(85.7)	8-60	23.3	< 3.6 - 34.1	
Wholesaler	Raw kernel	5	6.2 - 8.8 (7.5)	37.8-93.4 (63.6)	3.3-50.7 (29.0)	3.2-15.6 (7.4)	5	(100.0)	14-100	72.4	< 3.6-6065.9	
Retailer	Raw kernel	45	6.5 - 9.3 (8.5)	29.3-73.0 (56.3)	15.9-39.0 (29.0)	4.1-40.3 (14.7)	45	(100.0)	34-100	87.6	< 3.6-6073.0	



Moisture content of pods and kernels are closely related with the drying process. Sun-drying is the most critical postharvest handling procedure for peanuts, especially when the harvest coincides with the wet season. According to Wongvirajtana *et al.* (1993) the duration of sun-drying can significantly affect fungal growth in grains, with the longer the period of sun-drying, the more chance for the fungi to infect the grains.

Physical quality of kernels

Range and mean of physical quality characteristics of kernels derived from various kinds of peanuts collected from farmers, collectors, wholesalers and retailers are presented in Table 6, and Figure 2. The percentage of intact kernels of peanuts collected from farmers was the highest, followed by those collected from collectors, wholesalers and retailers. This was probably due to the peanuts which have not being shelled using mechanical or diesel powered sheller at the farmer level, while the duration of storage of peanuts was relatively short (< 7 days). The percentages of shriveled kernels in each of the delivery chain were relatively the same (more than 22%). SNI (1995) determined the maximum percentage of shriveled kernels is 4%, consequently the percentage of shriveled kernels in each delivery chain can be categorized as very high. This was probably due to the early harvest. Results of interviews with farmers showed that 59% of respondents harvest peanuts 90 days after planting (Table 2). Peanuts harvested before full maturity will tend to produce

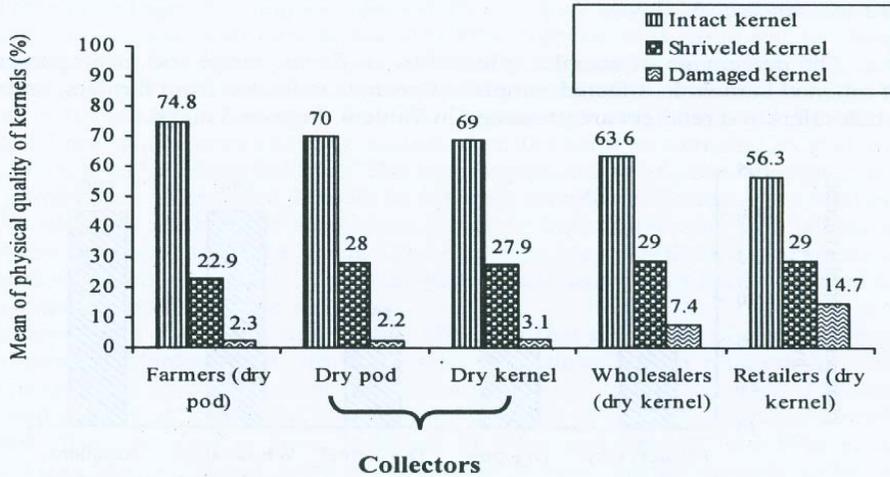


Figure 2. Mean percentages of physical quality of kernels. Peanut samples derived from various kinds of peanuts collected from different points of the delivery chain.

shriveled kernels after drying, consequently the kernels could be more easily infected by fungi.

The percentage of damaged kernels collected from retailers was the highest, followed by that collected from wholesalers, collectors and farmers. The damaged kernels could have been caused by insects, rodents and fungal attacks, and inappropriate equipment used for the shelling of pods. The highest percentage of damaged kernels at the retailer level was probably due to the longer duration of postharvest handling from farmer up to the retailer.

The range and mean percentages of intact kernels derived from various kinds of peanuts and collected from farmers, collectors, wholesalers and retailers were 59.6 - 83.9% and 74.8%; 42.2 - 83.0% and 70.0%; 56.2 - 77.3% and 69.0%; 37.8 - 93.4% and 63.6%; 29.3 - 73.0% and 56.3%, respectively. The range and mean-percentages of shriveled kernels derived from various kinds of peanuts and collected from farmers, collectors, wholesalers and retailers were 14.2 - 36.6% and 22.9%; 13.7 - 56.4% and 28.0%; 19.7 - 41.7% and 27.9%; 3.3 - 50.7% and 29.0%; 15.9 - 39.0% and 29.0%, respectively. The range and mean percentages of damaged kernels derived from various kinds of peanuts and collected from farmers, collectors, wholesalers and retailers were 0.6 - 4.9% and 2.3%; 0.6 - 5.6% and 2.2%; 2.1 - 4.5% and 3.1%; 3.2 - 15.6% and 7.4%; 4.1 - 40.3% and 14.7%, respectively.

The incidence of A. flavus

The percentage of samples infected by *A. flavus*, range and mean percentages of infected kernels in infected samples of peanuts collected from farmers, collectors, wholesalers and retailers are presented in Table 6, Figures 3 and 4.

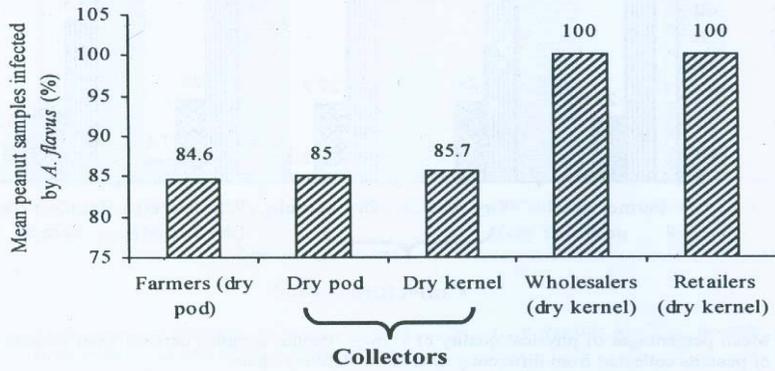


Figure 3. Mean percentages of peanut samples infected by *A. flavus*. Peanut samples derived from various kinds of peanuts collected from different points of the delivery chain.

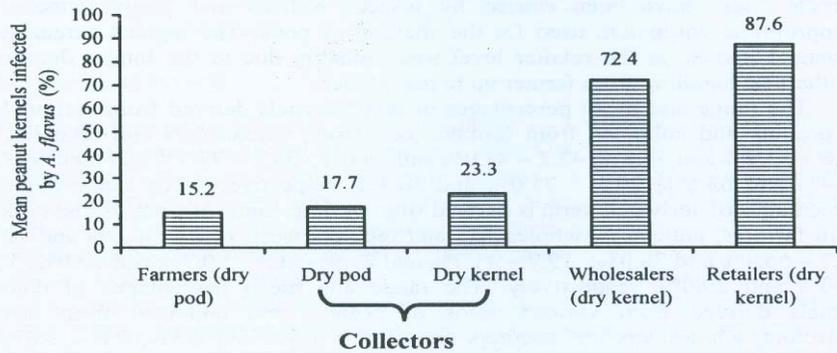


Figure 4. Mean percentages of kernels infected by *A. flavus* in infected samples. Peanut samples derived from various kinds of peanuts collected from different points of the delivery chain.

The percentage of samples infected by *A. flavus* was highest in peanuts collected from wholesalers and retailers (100%, respectively), followed by those collected from collectors in the form of pods (85%) and kernels (85.7%), and farmers (84.6%). The mean percentage of infected kernels in infected samples of peanuts collected from retailers was the highest (87.6%), followed by those collected from wholesalers (72.4%), collectors in the form of kernels (23.3%) and pods (17.7%), and farmers (15.2%). The highest percentage of peanut samples and mean percentages of infected kernels in infected samples collected from retailers was related to the methods of postharvest handling from farmers up to retailers, as well as the duration of storage. Peanut kernels were more easily infected by fungi compared to unshelled peanuts. The damaged kernels were more easily infected by fungi compared to intact kernels.

Dharmaputra and Retnowati (1996) observed that the range in percentage of peanut kernels infected by *A. flavus* in samples collected from retailers in some locations in West Java during the wet season was 83 - 100%. *Aspergillus flavus* was found in 98% of 256 peanut kernel samples and in 61% of all examined kernels collected from retailers in some locations in West and Central Java (Pitt *et al.* 1998). According to Dharmaputra *et al.* (2003) 24 raw kernel samples collected from retailers in traditional markets located in Bogor, Pati, Yogyakarta and Malang were 100% infected with *A. flavus* during both the wet and dry seasons, respectively. Dharmaputra *et al.* (2005) also reported that 98 and 100% of 54 raw peanut kernel samples collected from retailers during the wet and dry seasons in Wonogiri regency, West Java, were infected by *A. flavus*, respectively.

Aflatoxin B₁ contamination

The range of aflatoxin B₁ contents in peanut kernels derived from various kinds of peanuts and collected from farmers, collectors, wholesalers and retailers is presented in Table 6. The range of aflatoxin B₁ contents in peanut samples collected from retailers (< 3.6 - 6073.0 ppb) was the widest, but almost similar to those collected from wholesalers (< 3.6 - 6065.9 ppb).

The percentage of samples contaminated with different levels of aflatoxin B₁ is shown in Table 7. In Australia the maximum allowable limit of aflatoxin in peanut and peanut products is 15 ppb (QDPI 2000). The percentage of samples contaminated with more than 15 ppb of aflatoxin B₁ was highest in peanuts collected from wholesalers (80% of samples), followed by retailers (75.6%), farmers (38.5%) and collectors in the form of pods and kernels (30.0 and 14.3%, respectively), although the highest percentage of samples infected by *A. flavus* was found in peanuts collected from retailers, followed by those collected from wholesalers, collectors and farmers. This may have been associated with differences in the existence of toxigenic strains of *A. flavus*. According to Pitt and Hocking (1997) aflatoxin production depends on the toxigenicity of strains of *A. flavus*. The presence of antagonistic fungi to toxigenic *A. flavus* could also inhibit aflatoxin production. Dharmaputra *et al.* (2001) reported that *A. niger* was the most promising

antagonistic fungus to toxigenic strains of *A. flavus*, because it inhibited aflatoxin production up to 80% under *in vitro* conditions. The percentages of peanut samples infected by *A. niger* at farmer, collector, wholesaler and retailer levels in Cianjur regency were 69.2, 80 and 85.7, 60 and 76.5%, respectively.

Table 7. Number and percentage of peanut samples derived from various kinds of peanuts collected from different points of the delivery chain in Cianjur regency and the city of Cianjur (West Java) during the wet season (February 2004), and contaminated with different levels of aflatoxin B₁.

Aflatoxin B ₁ contents (range in ppb)	Number (%) of peanut samples contaminated with aflatoxin B ₁				
	Farmer – Dry pod	Collector		Wholesaler – Kernel	Retailer – Kernel
		Dry pod	Kernel		
≤ 5	5 (38.4)	15 (37.5)	5 (71.4)	1 (20.0)	10 (22.2)
> 5 ≤ 15	3 (23.1)	13 (32.5)	1 (14.3)	0	1 (2.2)
> 15 ≤ 50	3 (23.1)	3 (7.5)	1 (14.3)	0	4 (8.9)
> 50 ≤ 100	1 (7.7)	1 (2.5)	0	1 (20.0)	2 (4.4)
> 100 ≤ 500	1 (7.7)	4 (10.0)	0	0	16 (35.6)
> 500 ≤ 1000	0	3 (7.5)	0	0	5 (11.1)
> 1000	0	1 (2.5)	0	3 (60.0)	7 (15.6)

At the farmer level, no sample was contaminated with aflatoxin B₁ at more than 1000 ppb. At the collector, wholesaler and retailer levels, 1 sample (2.5% of collected samples), 3 samples (60% of collected samples) and 7 samples (15.6% of collected samples) contained more than 1000 ppb of aflatoxin B₁, respectively. At the collector level, aflatoxin B₁ content in one sample was 3000 ppb. The highest aflatoxin B₁ content measured at the wholesaler and retailer level was 6066 ppb (in one sample) and 6073 ppb (in one sample), respectively. Pitt and Hocking (1996) concluded that more than 1000 ppb of aflatoxin could cause acute toxic both in humans and animals.

Although the percentage of samples contaminated with aflatoxin B₁ exceeding 15 ppb at the farmer level (38.5%) was higher than that at the collector level (30%), aflatoxin B₁ contents more than 500 ppb were found in 4 peanut samples collected from collectors, while the highest aflatoxin B₁ content at farmer level was 114.2 ppb. This results showed that aflatoxin B₁ contents at the collector level are generally higher than at the farmer level, probably because the duration of storage at the collector level was longer, and under poorer storage conditions compared to that at the farmer level (Tables 2 and 3).

At the farmer level, 2 peanut samples were not infected by *A. flavus*, but they were contaminated with aflatoxin B₁ (Table 6). It was therefore assumed that these peanuts were infected by *A. flavus* before harvest, or during sun-drying, and contaminated with aflatoxin B₁ during sun-drying. Further sun-drying, and in the presence of antagonistic fungi to *A. flavus*, could inhibit or kill *A. flavus*. Therefore,

no *A. flavus* was isolated using AFPA media, but aflatoxin B₁ still existed. Buchi and Rae (1969) found that aflatoxin could only be degraded at 268 - 269°C.

CONCLUSIONS

In Cianjur regency farmers dried (sun-drying) peanut pods until safe moisture contents were obtained, and sold peanuts to collectors in the form of dry pods. In general, farmers stored peanuts in woven polypropylene bags for 1 - 7 days before selling to collectors. Collectors sold the peanuts to wholesalers and retailers in the form of dry pods and kernels. Collectors shelled the peanut pods using diesel powered shellers. At the wholesaler level, peanuts were stored in woven polypropylene and jute bags. In general, stacks of bags containing peanuts were not placed on pallets. Retailers stored peanuts in woven polypropylene and jute bags for 1 - 7 days. Most retailers used rectangular plastic basins as containers when the peanut sampling was being conducted, followed by winnowing trays, wooden boxes, round plastic basins, woven polypropylene bags and jute bags.

The mean moisture contents of peanut kernels derived from various kinds of peanuts in each part of the delivery chain were considered to have safe moisture contents. The percentages of intact peanut kernels collected from farmers was the highest, followed by those collected from collectors, wholesalers and retailers. The percentages of shriveled kernels collected from different parts of the delivery chains were relatively similar. The percentage of damaged kernels collected from retailers was the highest, followed by that collected from wholesalers, collectors and farmers.

The percentages of samples infected by *A. flavus* in peanuts collected from wholesalers and retailers were the highest (100%, respectively), followed by those collected from collectors and farmers. The mean percentages of infected kernels in infected samples collected from retailers were the highest, followed by those collected from wholesalers, collectors and farmers.

The range of aflatoxin B₁ contents in peanut samples collected from retailers (< 3.6 - 6073.0 ppb) was the widest, but almost similar with that collected from wholesalers (< 3.6 - 6065.9 ppb). The percentage of samples contaminated with more than 15 ppb of aflatoxin B₁ collected from wholesalers (80.0% of samples) was the highest, followed by retailers (75.6%), farmers (38.5%) and collectors in the form of pods and kernels (30.0 and 14.3%, respectively).

Our results show that postharvest handling methods employed prior to peanuts being delivered to wholesalers and retailers will have a severe impact on the level of aflatoxin contamination in peanuts in different parts of the delivery chain. Postharvest handling methods that minimize kernel infection by *A. flavus* and aflatoxin contamination, especially at wholesalers and retailers in traditional markets should be employed to minimize aflatoxin contamination for Indonesian peanut consumers.

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REFERENCES

- BSI. 1995. Oilseeds-determination of moisture and volatile matter content. British Standard International. (Supplement).
- Buchi, G. and I.D. Rae. 1969. The structure and chemistry of the aflatoxins. In Goldblatt, L.A. (Ed). Aflatoxins: Scientific Background, Control and Implications. Academic Press, New York. p. 55 - 75.
- Dhartnaputra, O.S and I. Retnowati. 1996. Fungi isolated from groundnuts in some locations of West Java. BIOTROPIA No. 9: 15 - 25.
- Dharmaputra, O.S., A.S.R.Putri, I. Retnowati and S. Ambarwati. 2001. Soil mycobiota of peanut fields in Wonogiri regency, Central Java: their effect on the growth and aflatoxin production of *Aspergillus flavus* *in vitro*. BIOTROPIA No. 17: 30 - 56.
- Dharmaputra, O.S., I. Retnowati, A.S.R. Putri and S. Ambarwati. 2003. *Aspergillus flavus* and aflatoxin in peanuts at various stages of the delivery chain in Pati regency, Central Java. Paper presented at the 3rd APEC/21st ASEAN Postharvest Technology Seminar. Nusa Dua, Bali, 23 - 26 August 2003.
- Dharmaputra, O.S., I. Retnowati, A.S.R.Putri and S. Ambarwati. 2005. *Aspergillus flavus* infection and aflatoxin contamination in peanuts at various stages of the delivery chain in Wonogiri regency, Central Java, Indonesia. Paper presented at the International Peanut Conference. Bangkok, 9-12 January 2005.
- Diener, U.L. and N.D. Davis. 1969. Aflatoxin formation by *Aspergillus flavus*. In Goldblatt, L.A. (Ed). Aflatoxins: Scientific Background, Control, and Implications. Academic Press, New York. p. 13-54.
- Lee, N.A. and I.R. Kennedy. 2002. ELISA Workshop Analysis of Aflatoxin B₁ in Peanuts. SEAMEO BIOTROP, Bogor, 12-13 February 2002.
- Lubulwa, A.S.G. and J.S. Davis. 1994. Estimating the social costs of the impacts of fungi and aflatoxins in maize and peanuts. In Highley, E., E.J. Wright, H.J. Banks and B.R. Champ. (Eds). Stored Product Protection. Vol. 2. Proceedings of 6th International Working Conference on Stored-product Protection. Canberra, 17-23 April 1994. CAB International, Wallingford. p. 1017 - 1042.

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- Pitt, J.I., A.D. Hocking and D.R. Glenn. 1983. An improved medium for the detection of *Aspergillus flavus* and *A. parasiticus*. *Journal Appl. Bacteriology* 54: 109-114.
- Pitt, J.I. and A.D. Hocking. 1996. Current knowledge of fungi and mycotoxins associated with food commodities in Southeast Asia. In Highley, E. and G.I. Johnson. (Eds). *Mycotoxin Contamination in Grains*. ACIAR Technical Reports 37, Canberra, p. 5 - 10.
- Pitt, J.I. and A.D. Hocking. 1997. *Fungi and Food Spoilage*. Blackie Academic and Professional, London.
- Pitt, J.I., A.D. Hocking, B.F. Miscamble, O.S. Dharmaputra, K.R. Kuswanto, E.S. Rahayu and Sardjono. 1998. The mycoflora of food commodities from Indonesia. *Journal of Food Mycology* 1 (1): 41 -60.
- QDPI. 2000. Aflatoxin in Peanuts; Tips to Reduce the Risk. Crop Link. Queensland Department of Primary Industries and Fisheries, Farming Systems Institute, Kingaroy. (See www.qld.gov.au/fieldcrops/3027.html).
- Standar Nasional Indonesia (SNI). 01-39219-1995. 1995. Kacang Tanah. Dewan Standardisasi Nasional, Jakarta.
- Wongvi raj tana, P., S. Soponronnarit and A. Nathakaranakule. 1993. Feasibility study of in-store corn drying under tropical climates. In Naewbanij, J.O., A.A. Manilay and A.S. Frio (Eds). *Increasing Handling, Processing and Marketing Efficiency in the Grain Postharvest System*. Proceedings of the 16th ASEAN Seminar on Grain Postharvest Technology. Phuket, 24 - 26 August 1993. ASEAN Grain Postharvest Programme, Bangkok, p. 265 - 283.