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36 **EFFECTIVENESS OF INDIGENOUS ENDOMYCORRHIZAL BIOFERTILIZER**
37 **PROTOTYPE ON ORGANIC SALAK LEAVES AND FRUITS IN BALI**

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46 **ABSTRACT**

47 Organically cultivated salak (*Salacca zalacca*) on dry land has limited production in Bali.
48 Typically, fertilization is carried out using leaf litter or other organic fertilizers because the fertility
49 of the soil is low for salak plantations. The present study analyzed the effectiveness of an
50 indigenous endomycorrhizal biofertilizer on the nutrient and total carbohydrate content of salak
51 leaves and fruits. The study used a randomized block design with nine replicates. The treatment
52 consisted of three levels, i.e., fertilization with leaf litter only as the farmers' way and control (C),
53 fertilization with indigenous endomycorrhizae biofertilizer prototype (P), and combined fertilization
54 with leaf litter and indigenous endomycorrhizae biofertilizer prototype (PM). Spores of indigenous
55 endomycorrhizae from salak plantations were used for making a biofertilizer prototype. The results
56 showed that P and CP treatments: 1) significantly increase the chlorophyll and relative water
57 content of leaves, and the number and weight of fruits per tree, 2) improve fruit quality by
58 increasing sweetness and weight per fruit, and 3) have a positive effect on water uptake and nutrient
59 absorption as indicated by high N and P of leaf tissue and high carbohydrate content of leaves.

60
61 **Keywords:** biofertilizer, endomycorrhizae, organic, prototype, *Salacca zalacca*.
62

63 **INTRODUCTION**

64 The province of Bali has been a major tourist destination in Indonesia. Increasing the number
65 of domestic and international tourists leads to an increased demand for fruits, including organic
66 salak fruits. The salak cultivation in Bali has been carried out organically since the 1500 years and
67 is hugely beneficial to the farmers; however, the quantity, quality, and continuity of organic salak
68 fruit produced is low (Rai *et al.* 2014).

69 This low productivity can be attributed to the nature-dependency of the farmers, who do not
70 yet implement adequate agricultural practices. Typically, fertilization only uses leaf litter, and
71 irrigation relies only on rainfall. Thus, the fertilization of organic salak renders low soil fertility
72 thereby decreasing the plant productivity over time (Rai *et al.* 2014). Rai *et al.* (2010) found that the
73 nutrient content of N, P, and K in the leaf tissue of organic salak in Bali was low; also, the soil
74 fertility was low as indicated by the levels of C-organic and the content of N, P, and K in the soils.
75 In order to maintain the soil fertility and environmental sustainability for high organic salak
76 productivity, the cultivation should be with indigenous endomycorrhizae biofertilizer due to the

77 diversity of fungal species of indigenous endomycorrhizae in nature is very large (Baslam *et al.*
78 2011; Proborini 2013; Suamba *et al.* 2014; Sarah & Ibrar 2016; INVAM 2017).

79 Kim *et al.* (2017) established a close correlation between soil, plants, and endomycorrhizae.
80 Indigenous endomycorrhizae isolated from plants in certain locations, will be more effective if
81 applied directly to the target plant concerned where it is taken as compared to the endomycorrhizae
82 brought from outside (non-indigenous endomycorrhizae). The arbuscular mycorrhizal symbiosis
83 between plants and fungi exerts a positive impact on improving the soil structure, expanding the
84 water absorption areas, increasing the plant tolerance to biotic and abiotic stresses, increasing the
85 nutrient uptake, and increasing the plant growth and yield (Nikhat 2014; Olagunju *et al.* 2014; Soka
86 & Ritchie 2016). In addition, endomycorrhizae increases the plant tolerance to various biotic and
87 abiotic stresses including alkalinity and metal toxicity (Abbasi *et al.* 2015; Bitterlich *et al.* 2018).
88 The endomycorrhizae fungus which is also called arbuscular mycorrhizae is beneficial to the host
89 plants as it plays a role in increasing nutrient absorption through the endomycorrhizal structures that
90 form large root surface areas such that the plant roots can absorb the nutrients (Baslam *et al.* 2011;
91 Sasvari 2012; Brundrett & Tedersoo 2018). Endomycorrhizae play a role in increasing plant
92 resistance to root pathogen attack through antibiotics produced during symbiosis with plant roots
93 that weaken and also kill the pathogenic bacteria, viruses, and fungi (Brundrett 2017; Sasvari,
94 2012).

95 Furthermore, the diversity and activity of indigenous endomycorrhizae in plant roots are
96 mainly determined by the type of the host plant (Ningsih *et al.* 2013; Sadhana 2014),
97 environmental growth factors, such as climate and soil moisture/drought (Hernadi 2012; Quiroga *et al.*
98 2017; Mathimaran *et al.* 2017; Kehri *et al.* 2018), and the level of soil fertility (Tahat & Sijam
99 2012; Kavitha & Nelson 2013; Mo *et al.* 2016). Rai *et al.* (2019a) explored and identified
100 morphologically and genetically the indigenous endomycorrhizae from salak root areas in three
101 regencies in Bali: Karangasem, Gianyar, and Tabanan. Herein, we identified two genera of
102 indigenous endomycorrhizae in salak: *Glomus* and *Entrophospora*. The genus *Glomus* consisted of
103 three species (*Glomus cubens*, *Glomus custos*, and *Glomus indicum*) while genus *Entrophospora*
104 consisted of only one species (*Entrophospora_sp_SH197095.06FU*). After propagation, the spores
105 of these species were formulated into prototypes of biological fertilizers/biofertilizer with carrier
106 media zeolite, quartz sand, sea sand, and volcanic sand. The most effective in salak seedling is a
107 prototype composed of a mixture of 100 spores from *glomus* species in 500 g volcanic sand carrier
108 media. This phenomenon was shown by the optimal growth of salak seedling and the maximal
109 (100%) colonization endomycorrhizae at the roots (Rai *et al.*, 2019b). Positive test results on salak
110 seedling growth need to be followed up by evaluating the efficiency of the prototype of indigenous
111 endomycorrhizae biofertilizer on organic salak plantations in the field. Thus, the present study

112 aimed to determine the effect of prototype indigenous endomycorrhizae biofertilizers on organic
113 salak production and its effect on the nutrient and carbohydrate content of the leaves.

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MATERIALS AND METHODS

116 The study was conducted from February to December 2019 in organic salak farms owned by
117 farmers in Sibetan village, Bebandem district, and Karangasem regency. The study used 15-year-
118 old salak trees and a randomized block design with nine replicates. The fertilization consisted of
119 three levels as follows: fertilized with leaf litter as the farmers' way/control (C), fertilized with
120 prototype indigenous endomycorrhizae biofertilizer (P), and combination of leaf litter and prototype
121 indigenous endomycorrhizae biofertilizer (CP). Therefore, it was needed 27 trees of salak with a
122 total plot size of 216 m square.

123 The spores of indigenous endomycorrhizae were taken from salak roots in Sibetan village.
124 The isolation of the spores was carried out using a wet filtering technique, followed by
125 centrifugation as described previously (Brundrett 2017). These spores were propagated according to
126 the method described by Rai *et al.* (2018), wherein corn was used as a host plant with water stress
127 treatment, and topping/shoot cutting was carried out at the age of 4 weeks. Watering to field capacity
128 was carried out from seed planting to 1 week of age, followed by watering 50% of field capacity
129 until 4 weeks of age. After topping at the end of 4th week, watering was stopped so that the plant
130 experiences stress. Then, the plant was dismantled to harvest the spores from the multiplication at
131 the end of the 5th week. Subsequently, the resulting spores were placed on volcanic sand: 100
132 spores/500 g sand.

133 Fertilization was conducted by making a hole, 30 cm wide and 20 cm deep, around the tree at
134 a distance of 40 cm from the base of the tree. The salak trees in treatment C were fertilized with 4
135 kg leaf litter of salak/tree. The leaves were cut into 20-cm-long pieces, placed in a hole, and covered
136 with soil. On the other hand, the trees in the P treatment were fertilized with prototype indigenous
137 endomycorrhizae made from a mixture of 100 spores of three species of *Glomus* (*Glomus cubense*,
138 *Glomus Custos*, and *Glomus indicum*) in 500 g volcanic sand carrier media per plant. The prototype
139 of the biofertilizer was evenly spread around the tree and covered with soil. Interestingly, the CP
140 treatment was carried out by spreading a prototype of the biofertilizer made from a mixture of 100
141 spores of three species of *Glomus* in 500 g of volcanic sand carrier media per plant, followed by
142 spreading the salak leaf litter on it evenly around the tree.

143 The chlorophyll content and the relative water content (RWC) of the leaves, the percentage of
144 fruit set, the number and weight of fruits per tree, weight per fruit, fruit diameter, fruit sweetness
145 level, N and P content of the leaves, and total sugar, reducing sugar (R-sugar) and sucrose content
146 of the leaves, and root colonization by indigenous endomycorrhizae were recorded. The

147 colonization of roots by indigenous endomycorrhizae was observed by the slide method of
148 Giovannetti and Mosse (1980) using the following formula: percentage of colonized roots = the
149 number of colonized roots/the total number of roots observed \times 100%. The percentage of fruit set
150 was calculated as follows: the number of flowers developed into fruits/the number of flowers \times
151 100%. The leaf chlorophyll content measured three times by Chlorophyll Meter SPAD-502 in
152 April, June, and August, and the average was calculated. The total sugar was analyzed by anthrone
153 method, R-sugar by Nelson–Somogyi method, and the sucrose content was calculated by
154 subtracting the value of total sugar content from that of R-sugar, and multiplied by 0.95. The RWC
155 was measured from matured leaves. The mature leaves after cutting from the tree then immediately
156 wrapped in aluminium foil, stored in icebox, and transported to the laboratory for the analysis.
157 Thirty pieces of leaf samples (10 pieces from the tip of the leaf, 10 pieces from the middle, and 10
158 pieces from the bottom), with a diameter of 1 cm, were taken from leaf sheaths using a round punch
159 and weighed (W_1). These leaf samples were immersed in water and irradiated with 40 W
160 fluorescent light at room temperature for 5 h. Then, each piece of leaf sample was carefully dried
161 with paper towel, and weighed (W_2), followed by oven-drying at 70 °C for 24 h before estimating
162 W_3 . The value of RWC was calculated by the formula: $(W_1 - W_2) / (W_3 - W_2) \times 100\%$. The nutrient
163 content of N and P leaves was analyzed by Kjeldahl method and the P-availability by Olsen
164 method, while number and weight of fruits per tree were calculated cumulatively at the end of the
165 study. Moreover, the weight of each fruit was calculated as follows: the total weight of fruit per
166 tree/the number of fruits per tree.

167 These data were analyzed using analysis of variance with SPSS (Statistical Product and
168 Service Solution) software 26th version. If the F test showed a significant difference, to differentiate
169 the average value among treatments the least significant difference (LSD) test was performed.

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RESULTS AND DISCUSSION

172 The results of the analysis of variance showed that the fertilization treatment significantly
173 affected all the observed variables. The weight and number of fruits per tree (790.80 g and 16.81
174 fruits, respectively) in the treatment of indigenous endomycorrhizae biofertilizer prototype (100
175 spores/500 g volcanic sand carrier media) were significantly higher than the treatments by
176 fertilizing with leaf litter/control wherein the value of weight and number of fruits per tree were
177 653.81 g and 14.94 fruits, respectively; however, the same did not differ significantly as compared
178 to CP (790.14 g and 16.56 fruits) (Table 1). Also, the quality of salak fruit in P and CP treatments
179 was significantly improved as compared to that in C, as shown by the increased sweetness of the
180 fruit (Table 2) and the weight per fruit (Table 1). The high number and weight of fruit per tree, as
181 well as the increased fruit quality due to the administration of indigenous endomycorrhizae

182 biofertilizer prototype, elevated the N and P content of the leaves as compared to the control (Table
 183 2); also, the chlorophyll content and RWC of the leaves were increased (Table 1). Visually, the
 184 trees treated with C and CP had dark green and fresh leaves as compared to the control.

185

186 Table 1 Effect of prototype indigenous endomycorrhizae biofertilizers on the leaf chlorophyll
 187 content, RWC of leaves, number and weight of fruits per tree, weight per fruit, and fruit
 188 sweetness in salak plants

Treatments	Leaves chlorophyll content (SPAD)	Leaves relative water content (%)	Number of fruit per tree (unit)	Weight of fruit per tree (g)	Weight per fruit (g)	Fruit sweetness (% Brix)
C	68.82 b	55.03 b	14.94 b	653.81 b	44.09 a	12.89 b
P	76.83 a	65.37 a	16.81 a	790.80 a	47.61 a	13.72 a
CP	77.46 a	65.75 a	16.56 a	790.14 a	47.97 a	14.66 a
LSD 5%	0.67	0.98	1.59	96.34	7.27	0.51

189 Note: The numbers followed by the same letter in the same column did not show any significant
 190 difference in the LSD level of 5%.

191

192 Higher production and yield quality in P and CP treatments as compared to control (C) were
 193 related to high leaf chlorophyll content (Table 1) and the ability of salak tree to absorb nutrients and
 194 water (Table 2). The high P and N content of the leaves in P and CP treatment is significantly
 195 associated with root colonization as compared to the C treatment. Table 2 displayed that root
 196 colonization by indigenous endomycorrhizae in P and CP is 100%, but only 23.14% in C. The high
 197 root colonization in P and CP revealed a mutualism symbiosis between indigenous
 198 endomycorrhizae and salak trees, which in turn, increases the P and N content and the RWC of
 199 leaves. The endomycorrhizae infects the roots of plants but does not cause injury and effectuating
 200 mutual reciprocal processes. Host plants obtain nutrients from endomycorrhizae, while
 201 endomycorrhizae obtain carbohydrates or food from host plants (Hernadi *et al.* 2012; Zasvari *et al.*
 202 2012).

203 High nutrient content and RWC of leaves in the P and CP treatments increase the chlorophyll
 204 content, which improves the photosynthesis as indicated by elevated total sugar, R-sugar, and
 205 sucrose content of leaves in the P and CP as compared to that in C (Table 2). The present study
 206 showed that indigenous endomycorrhizae fungus isolated from root of salak trees is suitable to be
 207 used as a biofertilizer. The administration of indigenous endomycorrhizae results in the absorption
 208 of nutrients and water through the endomycorrhizae structure that enlarges the surface area of salak
 209 root such that the plant roots can absorb the nutrients and water (Baslam *et al.* 2011; Sasvari 2012;
 210 Brundrett & Tedersoo 2018, Bitterlich *et al.* 2018). According to Beltrano *et al.* (2013) and Sarah
 211 & Ibrar (2016), spore density and root colonization of host plants are largely determined by the
 212 compatibility of endomycorrhizae with host plants, environmental factors, and interactions between

213 endomycorrhizae and chemical compounds produced by host plants. Thus, a correlation between
 214 indigenous endomycorrhizae biofertilizers prototype and salak trees can be suspected, which
 215 increases the yield, the quality of the yield, and the physiological processes of salak trees. The
 216 results of this efficiency test are in accordance with those of previous studies wherein
 217 endomycorrhizal fungi biofertilizer can increase the growth, production and quality of pineapple
 218 yield (Nurhandayani *et al.* 2013), chili (Tanwar *et al.* 2013), teak (Proborini, 2013), tea (Nepaleon
 219 *et al.* 2012) and apple (Fediala *et al.* 2018). Endomycorrhizae play a role in increasing plant
 220 resistance to drought or lack of water in the dry season because the root of the plants possess
 221 mycelium that can reach water in the wider rhizosphere area of the soil and adsorb water despite
 222 limited availability (Sasvari 2012). Furthermore, it is involved in producing various growth
 223 regulators such as auxins, cytokinins, and gibberellins and vitamins that can increase the growth of
 224 plant organs and roots that do not rapidly age and hence can function effectively in the absorption
 225 of nutrients and other solutes (Baslam *et al.* 2011; Tanwar *et al.* 2013). The endomycorrhizae also
 226 improve the soil structure because the mycelium on the outside of the roots of plants (covering soil
 227 grains) produces polysaccharide gels that increase the stability of soil aggregates (Kruger 2011;
 228 Sasvari 2012; Sadhana 2014, Jansa *et al.* 2016; Kim *et al.* 2017).

229

230 Table 2 Effect of prototype indigenous endomycorrhizae biofertilizer application on the N and P
 231 nutrient, total sugar, R-sugar, and sucrose content of the leaves and root colonization on
 232 salak plants

Treatments	N content of leaves (%)	P content of leaves (%)	Total sugar content of leaves (%)	R-sugar content of leaves (%)	Sucrose content of leaves (%)	Root colonization (%)
C	0,50 a	0.23 b	21.35 b	1.96 b	18.91 b	23.14 b
P	0,52 a	0.26 a	23.48 a	3.35 a	21.41 a	100.00 a
CP	0,53 a	0.27 a	24.71 a	3.41 a	21.73 a	100.00 a
LSD 5%	0,03	0.01	0.51	0.22	0.79	3.16

233 Note: The numbers followed by the same letter in the same column did not show any significant
 234 difference in the LSD level of 5%.

235

236 Although C was not given indigenous endomycorrhizae biofertilizer, root colonization
 237 occurred albeit with significantly lower intention as compared to that in P and CP treatments. The
 238 data showed that indigenous endomycorrhizae were naturally present in salak root area; however,
 239 for a positive influence on the production and quality of salak, the population needs to be increased
 240 by giving indigenous endomycorrhizae biofertilizer for the optimal production of the plants. The
 241 indigenous endomycorrhizal fungi, also known as arbuscular mycorrhizal fungi, in nature is
 242 diversified (Baslam *et al.* 2011; Proborini 2013; Suamba *et al.* 2014), with a positive effect on the
 243 salak trees (Juliadewi *et al.* 2014). Indigenous endomycorrhizae is naturally associated with plant

244 roots without human intervention and has a high potential for extensive colonization because it
245 recognizes the host plant and has a higher tolerance to environmental conditions, such as high stress
246 (Sadhana 2014; Quiroga *et al.* 2017). Based on these characteristics, it can be speculated that
247 indigenous endomycorrhizae isolated from salak roots has great potential to be developed as
248 biofertilizer because it adapts to host plants to be fertilized (Sasvari 2012; Sadhana 2014; Hohmann
249 & Messmer 2017).

250

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CONCLUSION

252 The current prototype of indigenous endomycorrhizae biofertilizer increases the yield and
253 quality of the fruits in organic salak. Also, the nutrient level, the relative water content of the leaves,
254 and the photosynthesis process were improved.

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REFERENCES

- 263 Abbasi, Hisamuddin, Akhtar H, Sharf R. 2015. Vesicular arbuscular mycorrhizal (VAM) Fungi: a
264 tool for sustainable agriculture. *Amer J Plant Nutr Fert Techno* 5(2):40-49.
- 265 Baslam M, Garmendia I, Goicoechea N. 2011. Arbuscular mycorrhizal fungi (AMF) improved
266 growth and nutritional quality of greenhouse-grown lettuce. *J Agr Food Chem* 59(10):5504-
267 5515.
- 268 Beltrano J, Ruscitti M, Arango MC, Ronco M. 2013. Effects of arbuscular mycorrhiza inoculation
269 on plant growth, biological and physiological parameters and mineral nutrition in pepper
270 grown under different salinity and p levels. *J Soil Sci Plant Nutr* 13 (1):123-141.
- 271 Bitterlich M, Rouphael Y, Graefe J, Franken P. 2018. Arbuscular mycorrhizas: a promising
272 component of plant production systems provided favorable conditions for their growth.
273 *Front Plant Sci* 29:1329. doi: 10.3389/fpls.2018.01329.
- 274 Brundrett MC. 2017. Global diversity and importance of mycorrhizal and nonmycorrhizal plants.
275 In Tedersoo L, editor. *Biogeography of Mycorrhizal Symbiosis*. *Ecological Studies* Vol.
276 230. Switzerland: Springer International Publishing. p. 533-556.
277 <https://doi.org/10.1007/978-3-319-56363-3-21>.
- 278 Brundrett MC, Tedersoo L. 2018. Evolutionary history of mycorrhizal symbioses and global host
279 plant diversity. *New Phytot* 220:1108-1115.
- 280 Fediala W, Mosa AE, Paszt LS, Frac M, Trzcinski P, Treder W, Klamkowski K. 2018. The role of
281 biofertilizers in improving vegetative growth, yield and fruit quality of apple. *Hort Sci*
282 45(4):173-180. <https://doi:10.17221/101/2017-HORTSCI>.

- 283 Giovannetti M, Mosse B. 1980. An evaluation of techniques for measuring vesicular arbuscular
284 mycorrhizal infection in roots. *New Phytol.* 84(3):489-500. [https://doi:10.1111/j.1469-](https://doi:10.1111/j.1469-8137.1980.tb04556.x)
285 8137.1980.tb04556.x.
- 286 Hernadi P, Sasvari Z, Albrechtova J, Vosatka M, Posta K. 2012. Arbuscular mycorrhizal inoculants
287 increase yield of spice pepper and affects indigenous fungal community in the field. *Hort*
288 *Sci* 47(5):603-606.
- 289 Hohmann P, Messmer MM. 2017. Breeding for mycorrhizal symbiosis: focus on disease
290 resistance. *Euphytica* 213(113):1-11. DOI 10.1007/s10681-017-1900-x.
- 291 INVAM (International Culture Collection of Vesicular Arbuscular Mycorrhizal Fungi). 2017.
292 Species descriptions from reference cultures. [http://fungi.invam.wvu.edu/the-fungi/species-](http://fungi.invam.wvu.edu/the-fungi/species-descriptions.html)
293 [descriptions.html](http://fungi.invam.wvu.edu/the-fungi/species-descriptions.html). (26 June 2017).
- 294 Jansa J, Smith AF, Smith SE. 2016. Are there benefits of simultaneous root colonization by
295 different arbuscular mycorrhizal fungi?. *New Phytol* 177:779-789.
- 296 Jha S, Kumar KN. 2011. Potential of mycorrhizal fungi in ecosystem: a review. *Inter J Res Bot*
297 1(1):1-7
- 298 Juliadewi KAC, Rai IN, Kartini NL. 2014. Aplikasi dosis jamur endomikoriza untuk meningkatkan
299 produksi buah salak Gula Pasir [Application of endomycorrhizal dosage for increasing
300 production of salak cv. Gula Pasir]. *Plumula* 4(3):19-25
- 301 Kavitha T, Nelson R. 2013. Diversity of arbuskular mycorrhizal fungi (AMF) in the rhizosphere of
302 *Helianthus annuus* L. *Amer-Eurasian J Agric & Environ Sci* 13 (7):982-987.
- 303 Kehri HK, Akhtar O, Zoomi I, Pandey D. 2018. Arbuscular mycorrhizal fungi: taxonomy and
304 its systematics. *Inter J Life Sci Res* 6(4):58-71.
- 305 Kim SJ, Eo J, Lee E, Park H, Eom A. 2017. Effects of arbuscular mycorrhizal fungi and soil
306 conditions on crop plant growth. *Mycobiology* 45(1):20-24.
- 307 Kruger M. 2011. Molecular phylogeny, taxonomy and evolution of arbuscular mycorrhizal fungi.
308 DNA-based characterization and identification [Dissertation]. Kumulative Dissertation der
309 Fakultat für Biologie an der Ludwig-Maximilians-Universität München Morgantown, West
310 Virginia Agriculture and Forestry Experimental Station.
- 311 Mathimaran N, Sharma MP, Raju MB, Bagyaraj DJ. 2017. Mycosphere essay 17 arbuscular
312 mycorrhizal symbiosis and drought tolerance in crop plants. *Mycosphere* 8(3):361-376.
313 Doi 10.5943/mycosphere/8/3/2.
- 314 Mo Y, Wang Y, Yang R, Zheng J, Liu C, Li H, Ma J, Zhang Y, C. Wei, Zhang X. 2016. Regulation
315 of plant growth, photosynthesis, antioxidation and osmosis by an arbuscular mycorrhizal
316 fungus in watermelon seedlings under well-watered and drought conditions. *Front Plant Sci*
317 7:644. Doi: 10.3389/fpls.2016.00644
- 318 Nepolean P, Jayanthi R, Pallavi RW, Balamurugan A, Kuberan T, Beulah T, Premkumar R. 2012.
319 Role of biofertilizers in increasing tea productivity. *Asian Pac J Trop Biomed* 13(2):1443-
320 1445.
- 321 Nikhat N. 2014. Potential of arbuscular mycorrhizal (AM) fungi in reclamation of waste lands. *Life*
322 *Sci* 2(3):48-50.
- 323 Ningsih DR, Kramadibrata K, Gunawan AW. 2013. Arbuscular mycorrhizal fungi associated with
324 bisbul (*Diospyros blancoi*). *BIOTROPIA* 20(2):112-121. DOI: 10.11598/btb.2013.20.2.2.
- 325 Nurhandayani R, Linda R, Khotimah S. 2013. Inventarisasi jamur mikoriza vesikular arbuskular
326 dari rhizosfer tanah gambut tanaman nanas (*Ananas comosus* L. Merr) [Inventory of

- 327 arbuscular vesicular mycorrhizal fungus from the peat rhizosphere of the pineapple (*Ananas*
328 *comosus* L. Merr) plant]. *J Protobiont* 2 (3):146-151.
- 329 Olagunju EO, Owolabi KT, Alaje DO. 2014. Effect of mycorrhiza on plant growth. *J Environ Sci*
330 *Toxicol Food Technol* 8(1): 83-85.
- 331 Proborini MW, Sudana M, Suarna W, Ristiati P. 2013. Indigenous vesicular arbuscular mycorrhizal
332 (vam) fungi in cashew nut (*Anacardium occidentale* L.) plantation of north east Bali Island-
333 Indonesia. *J Biol Agric Healthcare* 3 (3) 114 -121.
- 334 Quiroga G, Erice G, Aroca R, Chaumont F, Ruiz-Lozano J.M. 2017. Enhanced drought stress
335 tolerance by the arbuscular mycorrhizal symbiosis in a drought-sensitive maize cultivar is
336 related to a broader and differential regulation of host plant aquaporins than in a drought-
337 tolerant cultivar. *Front Plant Sci* 8:1056. doi: 10.3389/fpls.2017.01056.
- 338 Rai IN, Semarajaya CGA, Wiraatmaja W. 2010. A Study on the flowering phenophysiology of
339 *Gula Pasir* snake fruit to prevent failure of fruit-set. *J Hort* 20(3):216-222.
- 340 Rai IN, Wiraatmaja IW, Semarajaya CGA, Alit Astiari NK. 2014. Application of drip irrigation
341 technology for producing fruit of salak 'Gula Pasir' (*Salacca Zalacca* var. *Gula Pasir*) off-
342 season on dry land. *J Degrad Min Land Manage* 2(1)219-222.
- 343 Rai IN, Suada K, Praborini M, Wiraatmaja IW. 2018. Spore propagation of indigenous
344 endomycorrhizae from several rooting areas of snake fruit on different soil water content.
345 *Inter J Biosci Biotech* 5(2):155-167.
- 346 Rai IN, Suada IK, Proborini MW, Wiraatmaja IW, Semenov M, Krasnov G. 2019a. Indigenous
347 endomycorrhizal fungi at salak (*Salacca zalacca*) plantations in Bali, Indonesia and their
348 colonization of the roots. *Biodiversitas* 20(8):2410-2416. DOI: 10.13057/biodiv/d200840.
- 349 Rai IN, Suada IK, Wiraatmaja IW. 2019b. Effectiveness of indigenous endomycorrhizal
350 biofertilizer prototype on salak (*Salacca zalacca*). Paper presented at The 2nd International
351 Conference on Science, Technology and Humanities (ICoSTH 2019). The Patra Bali Resort
352 & Villas Bali, 14-15 November 2019.
- 353 Sadhana, B. 2014. Arbuscular mycorrhizal fungi (AMF) as a biofertilizer: a review. *Int J Cur*
354 *Microbiol App Sci* 3(4):384-400.
- 355 Sarah S, Ibrar M. 2016. Effects of arbuscular mycorrhizal fungi on spores density and root
356 colonization of four hybrids of sunflower (*Helianthus annuus* L.) at different rock phosphate
357 levels. *Sarhad J Agric* 32(4):258-266.
- 358 Sasvari Z, Magurno1 F, Galanics D, Nhu Hang TT, Hong Ha TT, Luyen ND, Huong LM, Posta K.
359 2012. Isolation and identification of arbuscular mycorrhizal fungi from agricultural fields of
360 vietnam. *Amer J Plant Sci* 3:1796-1801.
- 361 Schubler A, Schwarzott D, Walker C. 2001. A new fungal phylum, the glomeromycota: phylogeny
362 and evolution. *Trop Eco* 44(2): 207-215.
- 363 Soka G, Ritchie M. 2016. Contributions of AM fungi and soil organic matter to plant productivity
364 in tropical savanna soils under different land uses. *Rhizosphere* 4(1):1-8.
- 365 Suamba IW, Wirawan IGP, Adiartayasa W. 2014. isolasi dan identifikasi fungi mikoriza arbuskular
366 (FMA) secara mikroskopis pada rizosfer tanaman jeruk (*Citrus* sp.) di Desa Kerta,
367 Kecamatan Payangan, Kabupaten Gianyar [Isolation and identification of arbuscular
368 mycorrhizal fungi microscopically in citrus plant rhizosphere (*Citrus* sp.) in Kerta Village,
369 Payangan District, Gianyar Regency. *E-J Agro Trop* 3(4):201-208.
- 370 Tahat MM, Sijam K. 2012. Mycorrhizal fungi and abiotic environmental conditions relationship.
371 *Res J Environ Sci* 25:431-440. [http://dx.doi.org/ 10.3923/rjes.2012](http://dx.doi.org/10.3923/rjes.2012).

372 Tanwar A, Aggarwal A, Kadian N, Gupta A. 2013. Arbuscular mycorrhizal inoculation and super
373 phosphate application influence plant growth and yield of capsicum annum. J Soil Sci Plant
374 Nutr 13(1):55-66.

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