

1 **ACCEPTED MANUSCRIPT**

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ACCEPTED MANUSCRIPT

## SEA RANCHING OF *Holothuria atra*: EFFECTS OF STOCKING DENSITY AND STOCKING TIME

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### ABSTRACT

Strong market demand and uncontrolled exploitation and/or the inadequate management of fisheries have caused many stocks of sea cucumbers to be overexploited. One suggested effort to overcome this problem is sea ranching. Stocking density is the most important consideration in sea cucumber rearing; therefore, this present work is aimed at elucidating the best stocking density for sea ranching of *Holothuria atra*. *H. atra* was taken from the Panjang Island, Jepara waters and reared in bottom cages in Teluk Awur waters, Jepara with a density of 30, 20, or 10 individuals per cage measuring 2 m × 2 m × 1.8 m (with bottom area of 4 m<sup>2</sup>). Stocking times of *H. atra* were at the initial time of cage installation, the second and the third months after installation. Bottom sediment characteristics (i.e., chlorophyll a, b, phaeophytin, and total carotene) of the sea cucumber habitat and water quality in the cages were measured monthly during the study. The results showed that growth of *H. atra* fluctuated; low stocking density yielded a higher weight gain than high stocking density did. The highest weight gain was present in the density of 10 individuals/cage in the second stocking month. The highest survival rate of *H. atra* was seen in the condition of 30 individuals/cage (93%) at the third stocking month, which means that these sea cucumbers were only reared for three months. The highest mortality occurred at a density of 20 individuals/cage with the survival rate being low (45%) at the first stocking time or in the fifth month of rearing. There was fission evidence among *H. atra* reared in the cages, resulting in smaller organisms. Among the water quality parameters, the concentration of chlorophyll a, b, phaeophytin, and carotene in the sediment fluctuated according to the time of sea cucumber rearing caused by their feeding and bioturbation. The study results suggested to stock *H. atra* at low density during the second stocking month to get higher growth.

**Keywords:** chlorophyll, fission, growth, sediment, survival

### INTRODUCTION

In recent decades, invertebrate fisheries have expanded in catch and value worldwide (Anderson *et al.*, 2011). One increasingly harvested marine invertebrate group is the sea cucumber. In Indonesia, sea cucumber is called “teripang”, “trepanng”, “timun laut”, or “gamat” (Hartati *et al.*, 2015). Strong market demand and uncontrolled exploitation and/or inadequate fisheries management have led to many marine species, such as sea cucumber stocks becoming heavily overfished (Ambariyanto, 2017). One option suggested to overcome this problem is sea ranching.

Sea ranching is essentially an effort to release/grow cultured or wild juveniles into natural habitat and harvested when they reach commercially optimal size (Purcell *et al.*, 2012). There are

62 some advantages of sea ranching, for example, inputs are nominally lower, as the processes  
63 between release and harvest are largely left to availability of food in the habitat, and the level of  
64 care to sea cucumber reared is minimum. But this practice is still able to yield a marketable sea  
65 cucumber size. However, to our knowledge, published studies on sea ranching of sea cucumber in  
66 Indonesia is very limited Attempt to culture sea cucumber in Indonesia is mainly grow out  
67 practices, especially for *H. scabra* (Pangkey *et al.*, 2012). Concerning with sea ranching of sea  
68 cucumber, there are similar studies had been done in several countries mainly for *H. scabra* such as  
69 in Philippines (Juinio-Meñez *et al.*, 2012), Australia (Bowman, 2012), and Papua New Guinea (Hair  
70 *et al.*, 2016). While studies for other species were done in Ecuador and Mexico for *Isostichopus*  
71 *fuscus* (Mercier *et al.*, 2012) and Portugal for *H. arguinensis* (Domínguez-Godino *et al.*, 2015),  
72 Therefore, to avoid the overexploitation of natural populations, further research on the sea ranching  
73 of sea cucumbers is necessary. Such study would also be able to provide knowledge to ensure a  
74 better understanding and to apply in marine conservation, population genetics, and connectivity  
75 patterns.

76 There is a new trend among fishermen in Indonesia that is cultivating sea cucumbers in sea  
77 pens. Sea ranching practices provide a way to restore damaged fisheries without having to formalize  
78 no-take zones or establish fishing rights for sea cucumbers. In the case of sand fish or *H. scabra*  
79 (Juinio-Meñez *et al.*, 2012, Hair *et al.*, 2016) said that sea ranching of sea cucumber give additional  
80 activity for fishers, i.e. rearing small sea cucumbers harvested from the wild in sea pens until they  
81 reach a marketable size. As has been gathered by Purcell *et al.* (2012a,b), regarding both the recent  
82 published data and unpublished data from mariculture programs on sea cucumber in the Indo-  
83 Pacific that provide hatchery production, the use of juveniles (for experimental, sea or pond  
84 farming, sea ranching, and/or stock enhancement reasons), and proponents information, very little  
85 came from Indonesia. This might be due to a lack of international publications on sea cucumber  
86 research in Indonesia. Moreover, in the case of conservation, no published work is available on the  
87 sea ranching of sea cucumbers in Indonesia. With the declining natural stock of sea cucumbers  
88 (Hartati *et al.* 2015), it is urgent to conduct works on sea cucumber sea ranching for the  
89 continuation of production and conservation. Here, we propose sea ranching for *H. atra* (local  
90 name: “teripang hitam”/loly fish) because they represent a source of protein for human consumption  
91 as well bioactive molecules for marine pharmaceuticals, are ecologically important for their  
92 sediment bioturbation and remineralization, and also demonstrate a specific reproduction scheme  
93 (i.e., asexual reproduction through natural fission). Stocking density is a very important factor that  
94 affects growth in aquaculture. Therefore, the objectives of this study were to determine the best  
95 stocking density and stocking time for sea cucumber *H. atra* ranching. Specifically, this paper will

96 discuss the effects of stocking density and stocking time on the performance of *H. atra* and the  
97 characteristics of the bottom sediment in the cages used.

98

99

## MATERIALS AND METHODS

100 *H. atra* (180 individuals) were taken from Panjang Island, Jepara waters with sizes of 100 g  
101 to 150 g. Bottom cages measuring 2 m × 2 m × 1.8 m were installed in Teluk Awur waters, Jepara  
102 with a geographic position of 06°37'43.8"S and 110°38'31.7"E. A previous study (Hartati *et al.*,  
103 2017a) determined that this area is a suitable location for cages of *H. atra*, as the habitat has a  
104 muddy sand substrate with sea grasses and seaweed.

105 The stocking densities applied were 30, 20, and 10 individuals/cage (equivalent to 7.5, 5,  
106 and 2.5 individuals/m<sup>2</sup>), with three different stocking time, i.e., when the cages were first installed,  
107 the second and the third months after the cages were installed. The stocking density is based on  
108 Lavitra *et al.* (2010a) for *H. scabra* and Xing *et al.* (2012) for *Apostichopus japonicas*, and  
109 assumed to be affected by behavioral performance due to competition for area and food. Sea  
110 cucumber weights were measured monthly to determine their performance and the number of sea  
111 cucumbers alive at each time point was counted to determine their survival rate.

112 Samples of bottom sediment were taken monthly to measure the biomass of the micro  
113 phytobenthic organisms through their photosynthesis pigment (chlorophyll a, b, phaeophytin; and  
114 carotene). Biomass of microphytobenthos were measure as concentration of chlorophyll-a, -b  
115 according method of Jonge *et al.* (2012) and Kuczynska *et al.* (2015), phaeophytin (method of  
116 Montani *et al.*, 2012) and carotene (method of Androuin *et al.*, 2018). Water quality parameters  
117 (e.g., temperature, salinity, pH, and dissolved oxygen) were also measured using a water quality  
118 checker in situ.

119 The growth rate (weight gain) was calculated as follows (Xing *et al.*, 2012)::

$$120 \text{ Weight gain} = W_2 - W_1 \quad (1)$$

121  $W_1$  = weight of *H. atra* at  $T_{n-1}$  (grams)

122  $W_2$  = weight of *H. atra* at  $T_n$  (grams)

123 Survival rate was calculated as follows:

$$124 \text{ SR (\%)} = (N_t/N_0) \times 100\% \quad (2)$$

125  $N_t$  = Number of *H. atra* alive in the end of the experiment

126  $N_0$  = Number of *H. atra* stocked at the beginning of the experiment

127

128

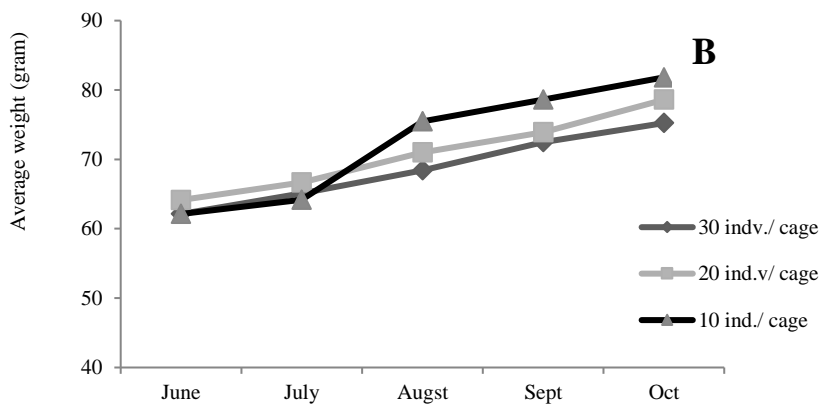
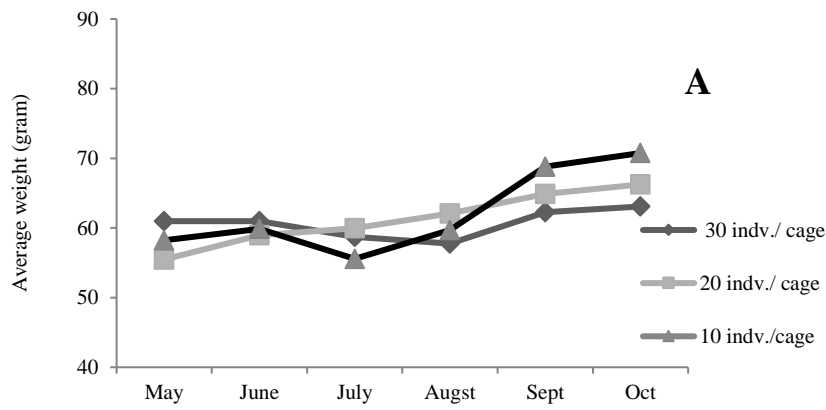
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## RESULTS AND DISCUSSION

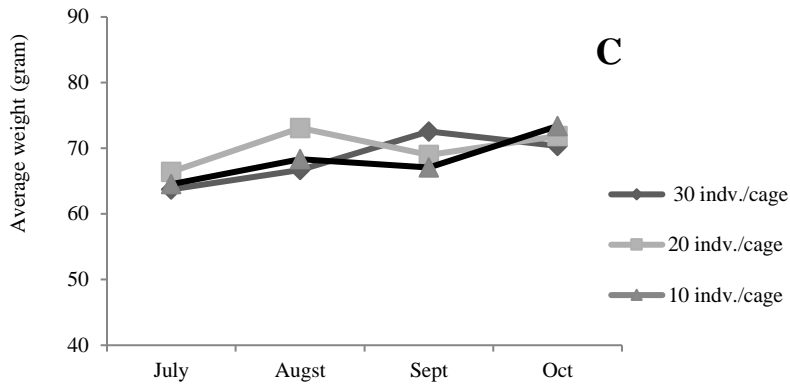
### The Growth and Survival Rates of *H. atra*

During the rearing period, the weights of *H. atra* were measured at the beginning of the study and every month thereafter until the study was completed. Increases in the weights of sea cucumbers were shown to be influenced by both the stocking density and time of stocking. Notably, a high stocking density affected the weight of the reared sea cucumber. Likewise, stocking on the second month after the bottom cage was installed was found to be higher weight gain than any of other time of stocking (Figure 1).

Sea cucumber growth was calculated based on the weight gain of *H. atra* at the end of the study and tended to fluctuate, in that low stocking density yielded a greater weight gain than did the higher stocking densities. The highest increase was obtained in the density of 10 individuals/cage at the second stocking. The lowest weight gain was seen with the stocking of 30 individuals/cage at the first stocking (Figure 2).

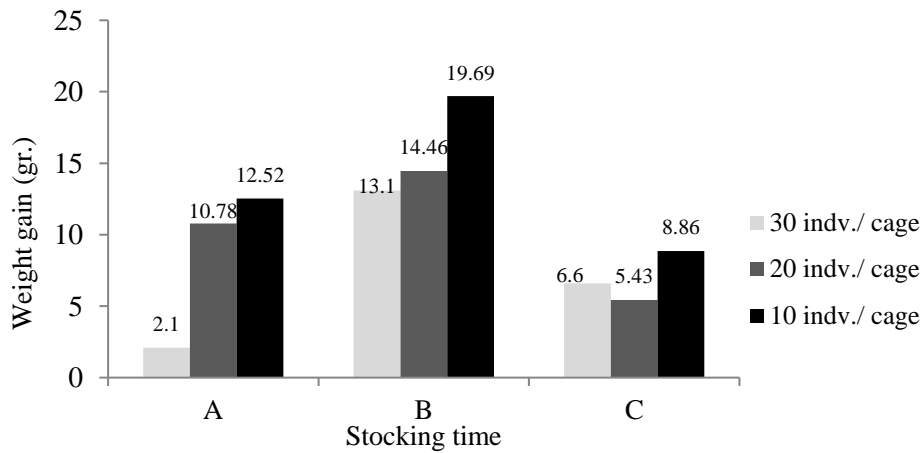


*Holothuria atra* stocking density and stocking time



146

147 Figure 1. Average weight of *H. atra* reared at different stocking times and densities (A = stocking at  
 148 Month 0 from the start of installation of the bottom cages, B = Month 1 after cages were  
 149 installed, and C = Month 2 after cages were installed).  
 150



151

152 Figure 2. Growth of weight of *H. atra* reared at different stocking times and densities (A = stocking  
 153 at Month 0 from the start of installation of the bottom cages, B = Month 1 after cages  
 154 were installed, and C = Month 2 after cages were installed).  
 155

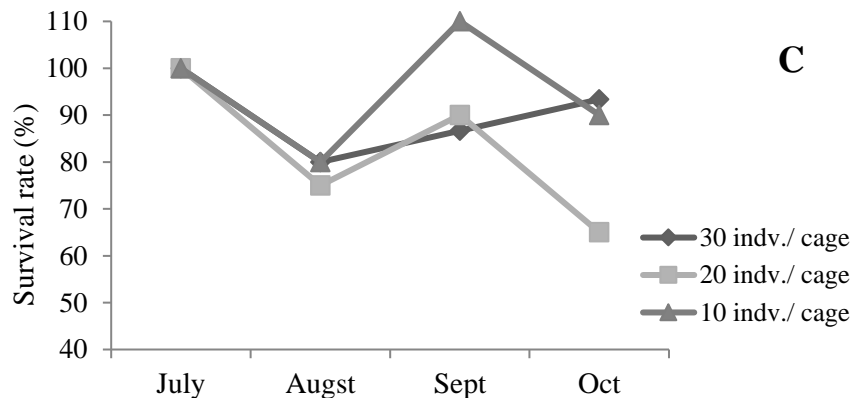
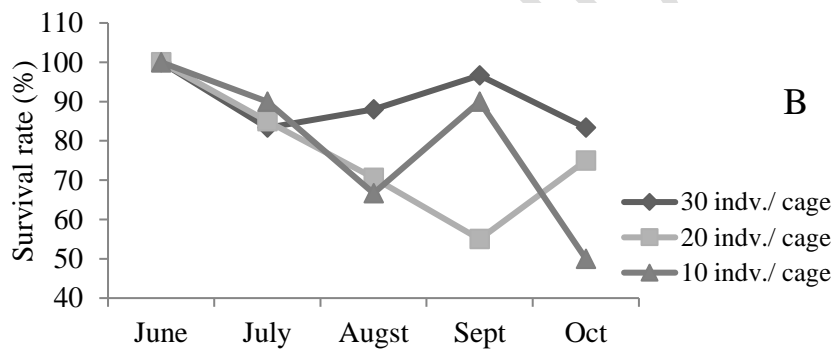
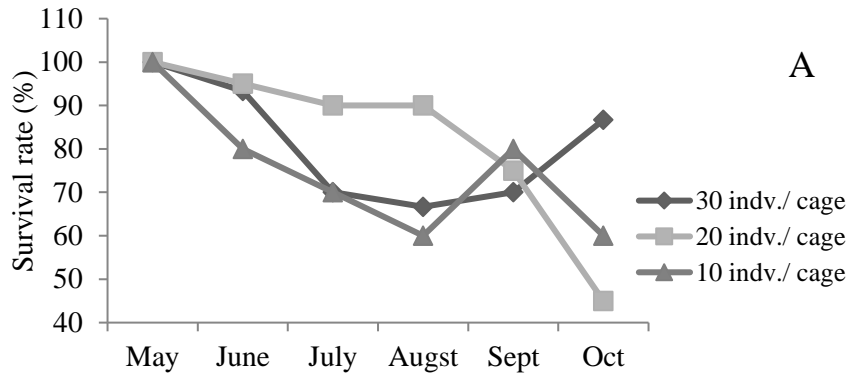
156 Stocking density is a very important factor that affects growth in aquaculture. At all times of  
 157 stocking, lower stocking density (i.e., 10 and 20 individuals/cage) culminated in a higher weight  
 158 than did higher stocking densities (e.g., 30 individuals/cage). The highest increase in weight (19.69  
 159 g) occurred with the stocking of 10 individuals/cage and the lowest (2.1 g) occurred with the  
 160 stocking of 30 individuals/cage. As noted in the study of Domínguez-Godino and González-  
 161 Wangüemert (2018), stress due to high stocking density is thought to be a major factor affecting the  
 162 growth and performance of the sea cucumbers *H. arguinensis* and *H. mammata*. Separately,  
 163 according to the results of research by Pei *et al.* (2012) on *Apostichopus japonicas*, stocking density  
 164 has an adverse effect on sea cucumber growth, not through decreasing water quality or food  
 165 competition but rather through the role of density as an environmental stress factor. The impact of  
 166 high density also leads to a greater variation in body weight, which is the same thing that happened  
 167 in this study.

168           The stress caused by crowding or overcrowding can stimulate the endocrine system of  
169 small-sized individuals, increasing cortisol levels in coelomic fluid and, in turn, accelerating energy  
170 consumption, modifying the energy budget model, and ultimately playing a negative role in the  
171 growth and composition biochemistry of these individuals (Pei *et al.* 2012). It seems that, in this  
172 study, the growth was also lowered by the effects of fission, which made the body size smaller. At  
173 high enough densities, sea cucumbers stop growing. The density value at which sea cucumbers can  
174 no longer grow is called the critical biomass value (CBV) (Lavitra *et al.* 2010a). The CBV for *H.*  
175 *scabra* is 650 g/m<sup>2</sup>, whereas that for juvenile *H. atra* is 250 g/m<sup>2</sup> to 350 g/m<sup>2</sup>. CBV is strongly  
176 influenced by the availability of food. *H. atra* is a deposit feeder (Asha *et al.*, 2015, Hartati *et al.*,  
177 2019b), so these organisms digest organic materials largely from sediments, which continue to  
178 decrease during maintenance. Sea cucumbers mainly eat microphytobenthos (algal epiphyte)  
179 (Paga'n-Jime'nez *et al.*, 2019) and bacteria (Lavitra 2010b) and the presence of sunlight reaching  
180 the bottom of the waters will support rapid growth of microphytobenthos which in turn will support  
181 growth of sea cucumbers (MacTavish *et al.*, 2012). The results of this study suggest to use the  
182 optimum stocking density to prevent problems such as overcrowding, which will cause low growth  
183 and/or variations in body size, deformation of the body shape, and the phenomenon "rotten  
184 stomach" (thickened stomach wall) (Seeruttun *et al.*, 2008). In this study, overcrowding caused  
185 fission, which in turn slowed the growth.

186           Stocking time in general also influenced the growth of *H. atra*: for first, second and third  
187 stocking periods, sea cucumbers were reared for five, four, and three months, respectively. Longer  
188 rearing (five months) led to less weight gain (2.1–12.52 g) versus the second stocking time (four  
189 months; 13.1–19.69 g). The second stocking provides an opportunity for sediments in the cages to  
190 be overgrown with members of the microphytobenthos such that the sediments are often more  
191 readily prepared for sea cucumber habitation as compared with at first stocking. At the third  
192 stocking, the maintenance duration of three months presented an average weight increase ranging  
193 from 5.43 g to 8.86 g.

194           The numbers of *H. atra* alive at the end of the study were different according to stocking  
195 density. Fluctuations in the number of sea cucumbers occurred, which, when calculated based on  
196 the survival rate, were remarkable in some cases due to exceeding 100% (occurring in the 10  
197 individuals/cage stocking density at the third stocking). This likely occurred because of the  
198 phenomenon of fission (asexual reproduction) in the sea cucumbers reared in cages. The survival  
199 trends of sea cucumbers stocked in dense stocking conditions and at different stocking times during  
200 maintenance are presented in Figure 3.

201 The highest survival rate of *H. atra* at the end of the study occurred in the 30  
202 individuals/cage stocking density, including 93% in the third stocking, which meant that sea  
203 cucumbers were only maintained for three months. The highest mortality occurred at the density of  
204 20 individuals/cage with the survival rate being 45% in the first stocking or in the fifth month of  
205 rearing. Detailed survival rate results are presented in Figure 4.

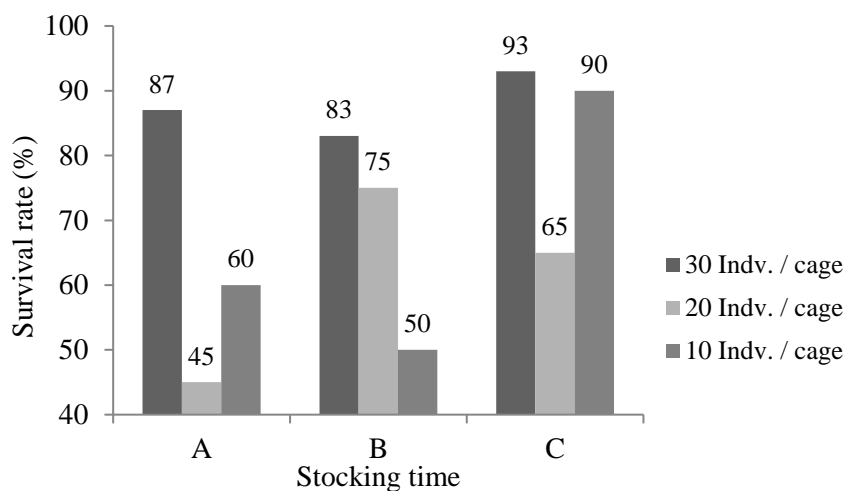


209 Figure 3. Survival rate of *H. atra* reared at different stocking times and densities (A = stocking at  
210 Month 0 from the start of installation of the bottom cages, B = Month 1 after cages were  
211 installed, and C = Month 2 after cages were installed).

214 The survival rates of sea cucumbers during rearing in cages fluctuated. Based on stocking  
215 density and stocking time, the survival rate at the 30 individuals/cage stocking density was higher



216 than other stocking densities. Based on stocking time, the third stocking still provided a high  
 217 survival rate because rearing took place for only three months. The high survival rate is thought to  
 218 be due to the existence of a fission process that increased the number of organisms living (Figure 3)  
 219 but resulted in smaller physical sizes (Figure 2).  
 220



221  
 222 Figure 4. Survival rate of *H. atra* reared at different stocking times and densities (A = stocking at  
 223 Month 0 from the start of installation of the bottom cages, B = Month 1 after cages were  
 224 installed, and C = Month 2 after cages were installed).  
 225

226 During being reared in cages with different stocking densities and stocking times, sea  
 227 cucumbers did participate in asexual reproduction or fission across all conditions (Table 1).  
 228 Although most of these phenomena were not directly observed, the organisms that are results of  
 229 fission can be distinguished from the original organisms (Figures 5). Sea cucumbers naturally have  
 230 a great potential to regenerate after the process of evisceration (expulsion of their internal organs).  
 231 Evisceration occurs due to physiological changes or in response to various external factors. During  
 232 evisceration, sea cucumbers secrete a large portion of their internal organs and, within a certain  
 233 period of time, such will grow back and they can continue to live normally (Hartati *et al.*, 2016).  
 234 Regeneration also occurs after the process of natural asexual reproduction by fission as well as  
 235 during fission stimulation (Hartati *et al.*, 2016). This process produces new individuals with  
 236 regeneration in the posterior or anterior part.

237  
 238 Table 1. Number of *H. atra* fissions during rearing in the cage

Stoking time	Stocking density		
	30	20	10
1	8	11	2
2	8	9	8
3	8	10	5

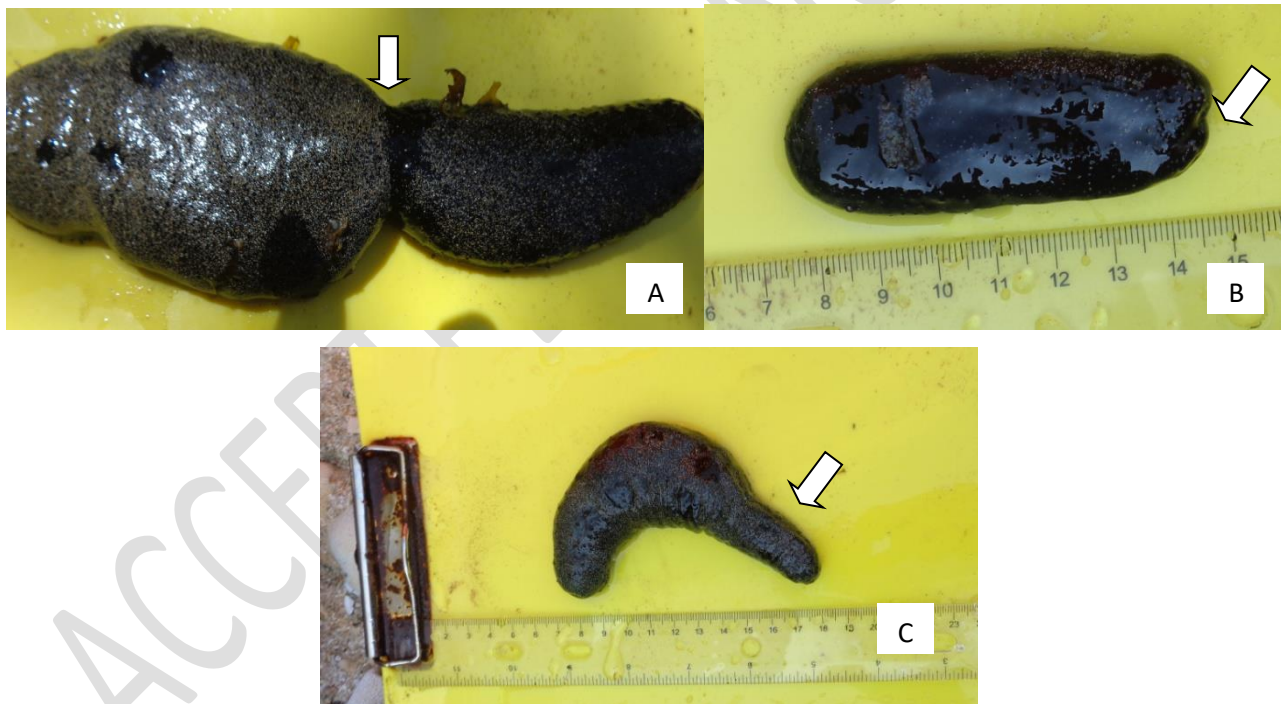
Total fissions	24	30	15
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239

240 The process of fission naturally occurs as presented in Figure 5; first, a narrowing of the  
241 part of the body that will be divided into two portions is observed (Figure 5A). After splitting  
242 (fission), a new individual closes the wound in the division section (fission plane) (Figure 5B) and  
243 the initial regeneration appears as a bulge in the fission plane. Then, regeneration of the body part  
244 happens, as shown in Figure 5C. Ultimately, this fission process contributed to high survival rates  
245 but produced smaller individual sea cucumber organisms.

246 In this study, in general, the more densely populated cages experienced more fission than  
247 did the nondense ones, but stocking time did not significantly affect the number of fission  
248 organisms (Table 1). According to Asha and Diwakar (2015), Asha *et al.*, 2015, Hartati *et al.*,  
249 (2019a), fission is a phenomenon that is highly dependent on population density,. In nature, fission  
250 is considered to play a role in maintaining populations in some types of holothurians to compensate  
251 for their mortality and migration (Dolmatov, 2014)).

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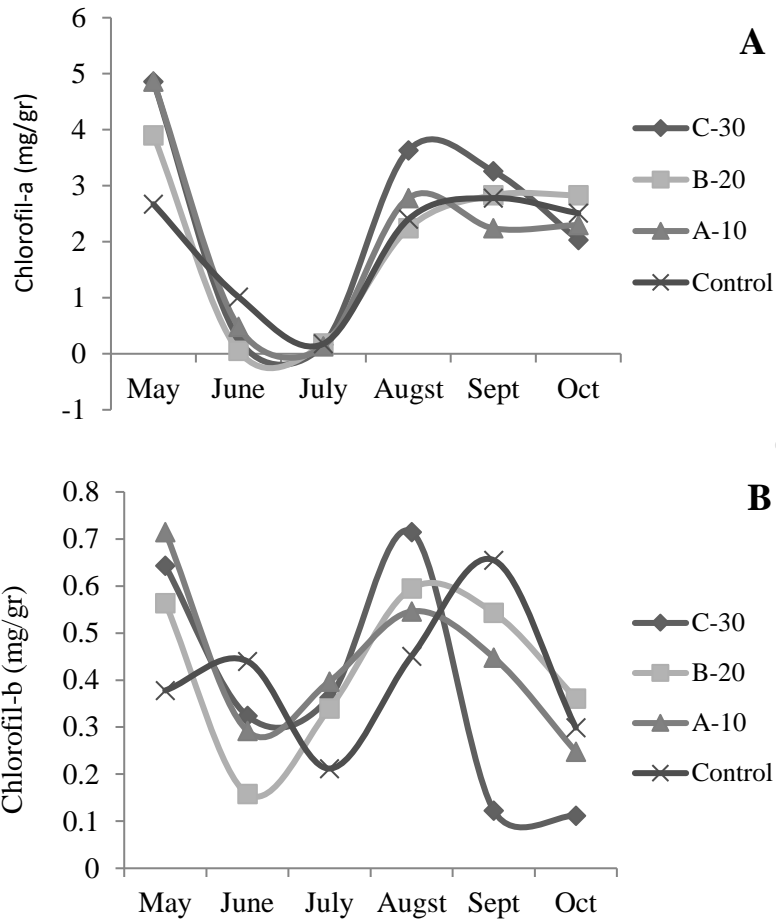
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254

255 Figure 5. Sea cucumbers that are undergoing fission (A); those that have just undergone fission,  
256 where the cleavage wound has been closed (B); and the regenerated missing part of the  
257 body (C) (→ arrow mark k is the position of fission/fission plane and regeneration).

258

259 Microphytobentic biomasses as food sources of *H. atra* were measured as concentration of  
260 chlorophyll a, b, phaeophytin, and total carotene levels in the sediment. Results are shown in  
261 Figures 6 and 7.



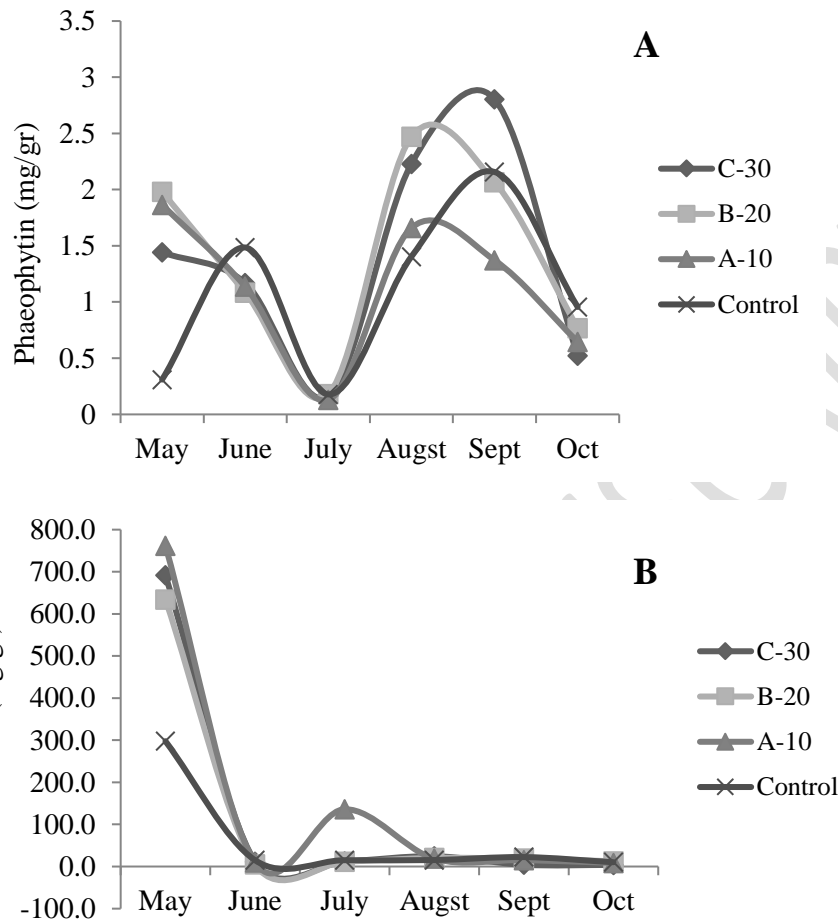
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263

264 Figure 6. Concentration of chlorophyll-a (A) and chlorophyll-b (B), in the sediment of *H. atra* cages  
 265 reared at different densities.  
 266

267 The trends in the concentrations of chlorophyll a, b, phaeophytin, and total carotene were  
 268 almost the same in all cages—specifically, high in the first month of rearing, decreased in the  
 269 second and third months of rearing, increased in the following (fourth) month, and then decreased  
 270 again in the last (fifth) month of the study (Figures 6 and 7). Microphytobenthic or benthic  
 271 microalgae are described as groups of photoautotrophic microorganisms that inhabit the surface  
 272 layers of sediment of shallow aquatic ecosystems such as diatoms, cyanobacteria, and other  
 273 chlorophytes (Paga'n-Jime'nez *et al.*, 2019). In shallow coastal waters, the microphytobenthos  
 274 plays an important role in the metabolic system by significantly contributing to primary producer  
 275 (Hardisoet *al.* 2013). Furthermore, the microphytobenthos includes important food sources for  
 276 meiofauna such as sea cucumbers (Mfilinge and Tsuchiya, 2016, Hartati *et al.*, 2017, Paga'n-  
 277 Jime'nez *et al.*, 2019). Holothurians can assimilate low content of organic matter as live diatoms,  
 278 bacteria and detritus by passing sediment through their gut system (Tolon *et al.*, 2015). This  
 279 consumption can cause fluctuations in concentrations of microphytobenthos, as stated by Hartati *et*  
 280 *al.* (2019b). The biomass of microphytobenthos could be measured as chlorophyll content in the

281 sediment (Du *et al.*, 2017) and the ratio of total chlorophyll c and b to chlorophyll a can be a good  
282 indicator of the amount of diatomic and green algal biomass (Cartaxana *et al.*, 2016)  
283



284

285

286 Figure 7. Concentration of phaeophytin (A) and total carotene (B) in the sediment of sea cucumber  
287 *H. atra* cages reared at different densities.  
288

289 According to Slater and Jeffs (2010), bottom sediment characteristics constitute one of the  
290 important components that influence sea cucumber habitat preferences and, hence, studies of the  
291 characteristics of sea cucumber sediment are very important to complete. Rocky sediment should be  
292 avoided because sea cucumbers live in mud or sand and remain in the organic material in the same  
293 place. *H. atra* is a deposit feeder that swallows sediment along with organic matter (Robinson *et al.*  
294 2013, Asha *et al.*, 2015), and the microbial composition in the intestine of adult holothurians  
295 have shown that they have a great diversity of microorganisms, such as bacteria, viruses, protozoa,  
296 and fungi that colonize their intestine (Paga'n-Jime'nez *et al.*, 2019).

297 Some studies have also shown that sea cucumbers swallow marine animal feces, including  
298 even their own feces (Barrio and. Tuya, 2013). Dissanayake and Stefansson (2012) stated that, in  
299 shallow water (<10 m) *H. atra* prefers sea grass habitats with sediments with 15% to 25% gravel  
300 and coarse sand (0.7–1.2 mm) but not mud. Separately, Hartati *et al.* (2017a) found that *H. atra* was

301 mainly present in sand-dominated habitats. The preference for specific habitat characteristics seems  
302 to be related to the needs of *H. atra* regarding feeding and protection (Hartati *et al.* 2017a,b).

303 Water quality parameters (e.g., temperature, salinity, pH, and dissolved oxygen) during  
304 the rearing of *H. atra* were measured and are presented in Figure 8. Water temperature is considered  
305 to be one of the most important parameters affecting the level of growth and development of sea  
306 cucumbers as well as their distribution in the sea. Temperatures in seawaters tend to be stable  
307 throughout the study period at 28 to 30°C (Figure 8). The temperature between cages were not  
308 significantly different because the cages were close to one another. According to Buccheri *et al.*  
309 (2019) because *H. atra* is tropical species, they perform optimally in the higher temperature  
310 environments in between 23-31°C. The process of aestivation (adaptation to high temperatures) has  
311 not been widely discussed in *H. atra*, but, usually during the day, *H. atra* will cover the body with  
312 sand, which is thought to be a response to higher temperatures. (Hartati *et al.*, 2019a).

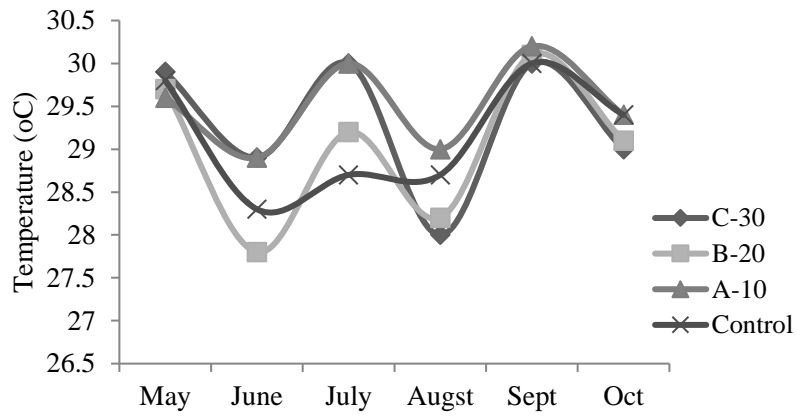
313 Sea cucumbers are stenohaline invertebrates that are unable to tolerate a wide range of  
314 salinity. Therefore, their placement in river mouths, estuaries, and other bays where salinity falls  
315 below 10 ppt during the monsoon season should be avoided. Although there is a small river that  
316 goes into the Teluk Awur waters, it did not affect the salinity around the cages, which were located  
317 500 meters from river mouth. Salinity levels in the Teluk Awur waters are presented in Figure 8,  
318 which ranges from 27.5 to 30.5‰, which is still suitable for the life of sea cucumbers.

319 In addition to temperature and salinity, water pH can affect the life of *H. atra*. pH  
320 fluctuations occur seasonally, in that the dry season is characterized by high water temperatures and  
321 low water movements/currents that produce low pH values between 7.3 and 7.5, while pH increases  
322 to between 8.4 and 8.6 during the rainy season because of the entry of high water influx (Gullian  
323 and Preciat, 2017). Wittmann and Portner (2013) said that, in general, vertebrates have the ability to  
324 regulate osmotic pressure and very good acid–base conditions and therefore are better able to cope  
325 with changes in pH as compared with invertebrates, which have a lower regulatory capacity, but  
326 information about the effects of pH on sea cucumber is limited.

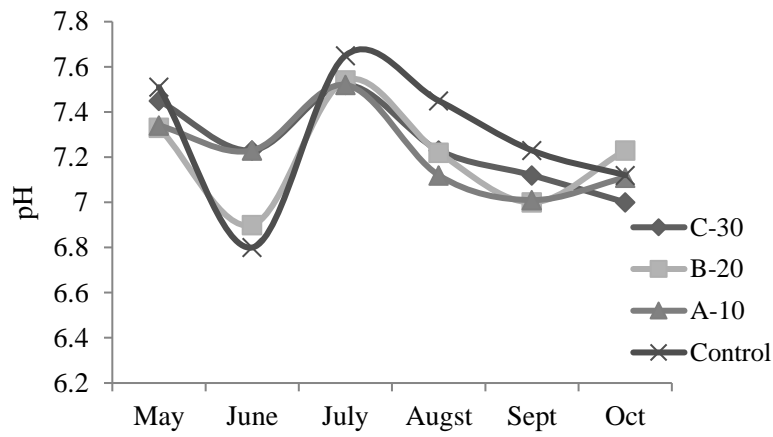
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*Holothuria atra* stocking density and stocking time

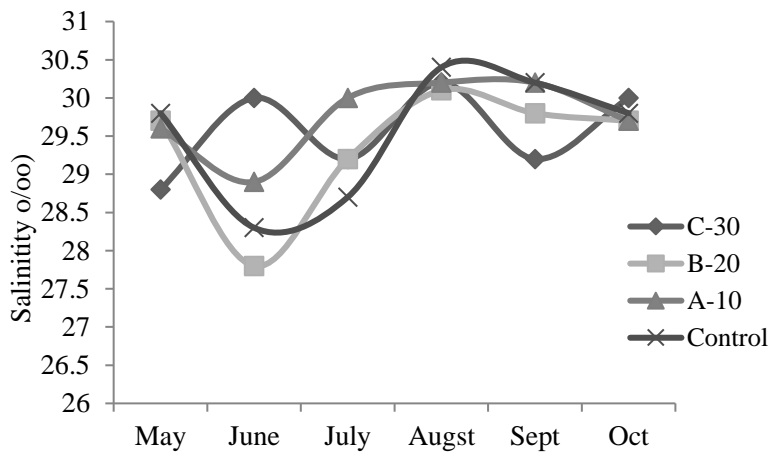
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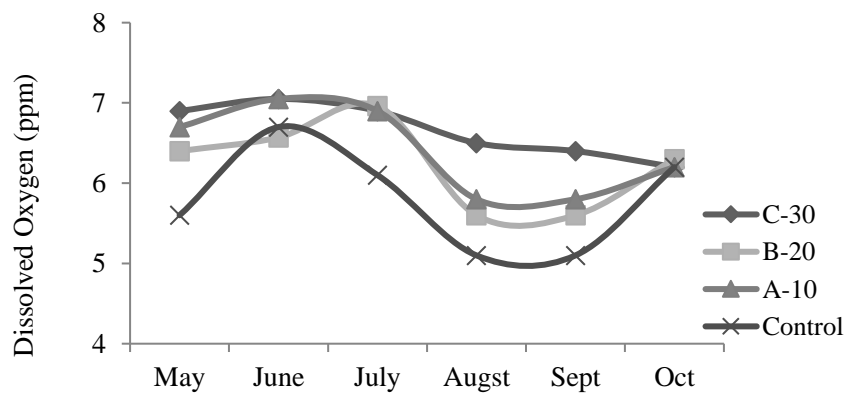
329



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331



332 Figure 8. Average temperature, salinity, pH, and dissolved oxygen of the sea water in *H. atra*  
 333 rearing cages with different stocking densities.

334

335           Sea cucumbers respire with their tentacles, skin, and respiration trees by consuming a  
336 significant amount of water and absorbing the dissolved oxygen inside. In their research, Zamani *et*  
337 *al.* (2018) found that the limit of hypoxia for *H. leucospilota* was 3 mg O<sub>2</sub>/L, wherein the sea  
338 cucumber released its cuverian tube to lower its energy needs in its aesthetic process. Furthermore,  
339 it has also been said that the normal conditions of dissolved oxygen levels of > 6 mg/L provide a  
340 100% survival rate. In the maintenance of *A. japonicas*, dissolved oxygen is maintained at > 5 mg/L  
341 (Xie *et al.* 2013). In this study, during the rearing period, the dissolved oxygen content in the cages  
342 ranged from 6 mg/L to 7 mg/L (Figure 8) and there were no signs of *H. atra* evisceration due to  
343 hypoxia.

344           The rearing of sea cucumbers in bottom cages has been carried out previously by Purcell  
345 and Agudo (2013), Hartati *et al.*, (2016) for both *H. scabra* and *Stichopus hermannii* with no food  
346 addition. Although according to Yokohama (2013) and Mahmoud *et al.* (2017) the maintenance of  
347 sea cucumbers with feeding greatly affects growth performance (i.e., sea cucumbers grow faster  
348 when given additional feed), in the process of rearing sea cucumbers in this study, the *H. atra* were  
349 not given additional feed as it was thought that the sediment/substrate would provide ample feed for  
350 sea cucumbers. On the subject of sea cucumber maintenance with the aim of sea ranching, no input  
351 is given, so, in current practices, the life of sea cucumbers is left to/depends largely on the  
352 availability of natural feed (Purcell *et al.*, 2012). Therefore, choosing the right location in this regard  
353 is very important to consider.

354

355

## CONCLUSION

356           *H. atra* reared in the bottom cages in the Teluk Awur waters of Jepara with low stocking  
357 densities (10 individuals/cage) presented higher growth and survival rates. Stocking time in the  
358 second month after the cages were installed also resulted in higher growth, as it allowed the food  
359 organisms available for the sea cucumber to be developed. Chlorophyll a, b, phaeophytin, and  
360 carotene levels fluctuated according to the time of sea cucumber rearing caused by their feeding and  
361 bioturbation activities.

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363

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