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ACCEPTED MANUSCRIPT

17 **THE INCREASING LEVEL OF INTERLEUKIN IN THE *Zingiber cassumunar*-TREATED**
18 **MICE**

19
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24

25 **ABSTRACT**

26 The cytokine is one of the proteins responsible for the immune system. Several types of
27 cytokines acting as key regulators of infection include IL-10, IL-12, and IL-14. The chemical content
28 of *Zingiber cassumunar* shows potential immunomodulatory effects. This study aimed to determine
29 the effect of the ethanol extract of *Zingiber cassumunar* (EEZC) on the expressions of IL-10, IL-12,
30 and IL-14. The test animals were BALB/c mice, which were divided into five groups, i.e., normal
31 group (untreated), negative control group (treated with 10% of tween 80), and three treatment groups
32 that respectively received 1.25 mg, 2.5mg, and 5mg/20g BW of EEZC. The treatment was carried out
33 for 21 days. On the 22nd day, the mice were induced with LPS intraperitoneally (except for the normal
34 group). The interleukin expression was observed by immunohistochemistry using specific antibodies,
35 and the expressed cells were counted under a microscope. The administration of EEZC at the doses
36 of 1.25 mg, 2.5mg, and 5mg/20g BW for 21 days increased the expression of IL-10, IL-12, and IL-
37 14 significantly and proportionally to the dose. and suggested the potency of extract to induce both
38 innate and adaptive immunity. This activity may be attributable to curcumin as an active compound
39 in this extract.
40

41 **Keywords:** curcumin, immunomodulator, interleukin, *Zingiber cassumunar*
42

43 **INTRODUCTION**

44 The immune system is responsible for protecting the host from various pathogenic
45 microorganisms. At the same time, it functions as a control of immune responses and prevents over-
46 reaction to the body's own cells (Saraiva and O'Garra, 2010). A decrease in the immune system can
47 affect the body's strength to fight infections or other diseases. Therefore, the presence of
48 immunomodulator compound that can improve the immune response to disease or infection is
49 necessary.

50 Immunomodulator could be defined as a substance, which can stimulate, suppress or modulate
51 any of the components of the immuno system including both specific and nonspecific immune system
52 (Das et al., 2014). Modulation of the immune system was remarked by induction, expression,
53 amplification or inhibition of certain part in the immune signaling and response. Thus,
54 immunomodulator is a substance used for its effect on the immune system.

55 Interleukin-10 (IL-10) is an anti-inflammatory cytokine that has a crucial role in preventing
56 inflammatory and autoimmune pathologies. It can both impede pathogen clearance and ameliorate
57 immunopathology. Many different types of cells can produce IL-10, with the major source of IL-10

58 varying in different tissues or during acute or chronic stages of the same infection (Couper et al.,
59 2008). IL-10 has a central role in infection by limiting the immune response to pathogens and thereby
60 preventing damage to the host. The production of IL-10 was associated with regulatory T (Treg) cells.
61 (Saraiva and O'Garra, 2010).

62 Interleukin-12 is an important cytokine in immunoregulation produced mainly by antigen-
63 presenting cells (APC), including macrophages and dendritic cells that respond to microbes (Abbas
64 et al., 2017). During the immune response, IL-12 is produced as a reaction to stimuli of various
65 compounds (including lipopolysaccharide/LPS).

66 Interleukin-14 (IL-14) is one of the cytokines produced by the immune system. It is produced
67 by activated B cells and T cells (Leca et al., 2008). The role of IL-14 is to regulate B-cell proliferation.
68 IL-14 can increase antibody responses to vaccinations causing autoimmunity and contribute to B-cell
69 lymphoma formation (Shen et al., 2006).

70 *Zingiber cassumunar*, known locally as *bengle* (Javanese, Indonesia), has been used for
71 treating various diseases traditionally. *Z. cassumunar* belongs to Zingiberaceae family and contains
72 terpenoids, essential oil, and curcuminoids. Several studies on *Z. cassumunar* reported some activities
73 including anticancer (Varalakshmi et al., 2008), antioxidant (Vankar et al., 2006) (Bua-in and
74 Paisooksantivatana, 2009) and immunomodulator (Nurkhasanah et al., 2017; Rahmawati, 2013). This
75 plant has been reported to exhibit an immunomodulatory activity by increasing phagocytic activity *in*
76 *vitro* (Chairul et al., 2009), increasing of nitric oxide and reactive oxygen species (Nurkhasanah et
77 al., 2017) and decreasing of malondialdehyde product in *Plasmodium berghei*-infected mice
78 (Nurmasari et al., 2014).

79 This study presents the activity of ethanol extract of *Zingiber cassumunar* (EEZC) in
80 stimulating the immune response *in vivo* as observed from its effect on interleukin-10, -12 and -14.
81 The present study will focus on these cytokines due to the important function of these cytokine in the
82 immune respons, either innate and adaptive immunity. The expression IL-10 and IL-14 are close
83 related to activation of adaptive immunity while the IL-12 is important in cell communication
84 between the macrophage and T cells. The study was carried on Balb/c male mice for 28 days treatment
85 and through oral administration. This *in vivo* study provides evidence of higher effectiveness of EEZC
86 in increasing immune response. This evidence was important information for development of *Z.*
87 *cassumunar* to be an immunomodulatory product.

88

89

MATERIALS AND METHODS

Materials

91 The *Zingiber cassumunar* rhizome was collected from a local market in Yogyakarta and
92 identified in the Biology Laboratory, Universitas Ahmad Dahlan. The test animal was obtained from

93 the Animal House of the Integrated Research and Testing Laboratory, Universitas Gadjah Mada
94 (LPPT UGM) Yogyakarta, Indonesia.

95

96 **Extraction**

97 The rhizome was selected, washed, and then sliced. The sliced rhizome was dried in an oven
98 at a temperature of 50°C. Afterward, the dried rhizome was blended or ground into powder. The
99 extraction was carried out by maceration method using 96% ethanol as the solvent. The maceration
100 lasted for 24 hours, and the yield were evaporated in a vacuum rotary evaporator to obtain a
101 concentrated extract. The concentrated extract was used for treatment and designated as EEZC.

102

103 **Thin-layer Chromatography (TLC) analysis of the extract**

104 The TLC analysis was carried out to identify the active compound of EEZC. A total of 100.0
105 mg of EEZC was dissolved in 10.0 ml of absolute ethanol. This procedure used curcuminoids (Sigma)
106 that was dissolved in ethanol as a standard. Each 2 µL of the extract and curcuminoid were applied
107 on silica gel GF 254 as the stationary phase and eluted with the mobile phase of chloroform: ethanol:
108 glacial acetic acid (94: 5: 1). The detection of active compound was done under daylight and UV 254
109 nm.

110

111 **Animal treatment**

112 The procedure of the study and the use of test animal were ethically approved by the Research
113 Ethics Committee of Ahmad Dahlan University on February 9, 2016, with Reference No. 011601011.

114 The test animals, i.e., BALB/c mice (8 weeks), were acclimatized for a week before the
115 treatment. The mice were divided into five (5) groups, namely normal group, negative control group
116 which treated with the solven (solution of tween 80 10%), and 3 treatment groups (1.25 mg/20gBW;
117 2.5 mg/20gBW; 5 mg/20gBW). The administration of EEZC was carried out orally for 21 days (3
118 weeks), once a day. On the 22nd day, the mice were sacrificed using CO₂ gas. Following sacrificing,
119 lipopolysachcharide (LPS) (Sigma) with dose 0.01 mg/20 g BW was injected into the peritoneal
120 cavity area, and after 1 hour, the macrophage was isolated and the expression of interleukin-10, -12
121 and -14 was observed with immunohistochemistry methode using specific antibody of IL-10, IL-12
122 and IL-14.

123

124 **Macrophage isolation**

125 Following the 21-day treatment, the mice were injected with LPS in the intraperitoneal cavity.
126 Then, the mice were dissected by opening the skin in the peritoneal area. As much as 10 ml of Roswell
127 Park Memorial Institute (RPMI) (Sigma) medium was injected into the stomach. The stomach was

128 massaged, then the RPMI medium was drawn again. The medium was centrifuged for 10 minutes,
129 and the supernatant was removed. The macrophage was washed with the medium and incubated for
130 24 hours. After overnight incubation, the macrophage was harvested.

131

132 **Immunohistochemistry assay**

133 The immunohistochemistry assay was based on the method reported in a previous study
134 (Nurkhasanah, 2015). The method was based on indirect method using specific primary antibody and
135 was conjugated with secondary antibody and chromogen. The interleukin expressed was observed as
136 brown colour as product of DAB (dimethyl amino benzidine) chromogen and detected under light
137 microscope.

138 The cultured macrophage was fixed with 1 ml of methanol. The methanol was removed, and
139 the macrophage was washed in PBS (phosphate buffer saline) for 5 minutes. The preparation was
140 then immersed in peroxidase blocking solution for 10 minutes at room temperature. Afterward, it was
141 washed under running water and then with PBS. A total of 50 μ l of blocking serum was added to the
142 preparation and then incubated in a humid temperature for 10-15 minutes. Then, 100 μ l (with dilution
143 1:100) of specific antibody (anti-IL-10, anti-IL-12, and anti-IL-14, murine recombinant, Biovision)
144 was added to the preparation and incubated on a moist tray at room temperature for 1 hour. After the
145 incubation, the preparations were washed with 1 mL of PBS. A total of 50 μ l (with dilution 1:100) of
146 antimouse biotin secondary antibody (Biovision) was added to each preparation, which was incubated
147 at a humid temperature for 20 minutes and then washed with PBS.

148 The preparations were incubated with 50 μ l of the streptavidin-peroxidase enzyme for 10
149 minutes, washed with PBS, and incubated with 50 μ l of DAB chromogen (peroxidase substrate
150 solution). Afterward, the preparations were washed with PBS and incubated with Mayer's
151 hematoxylin as the counterstain. The process was followed by washing with PBS. The macrophage
152 was ready for microscopic observation at 400x magnification. Macrophages that expressed
153 interleukin would show as brown stains.

154 The observation was carried out in some of field of view. The number of positive cell
155 expression was compared with the whole cell observed and presented as percentage value. The result
156 of treated group was compared with control group with statistical analysis to analyze the effect of
157 treatment.

158

159 **Statistical analysis**

160 The quantitative data of percentage expression of IL-10, IL-12 and IL-14 was analyze statistically for
161 normality and homogeneity. The analysis was followed with variance analysis using ANOVA and
162 followed by LSD for analysis between group of treated.

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RESULTS AND DISCUSSION

Extraction

The ethanol extract of *Zingiber cassumunar* rhizome was dark brown with a specific odor, thick consistency, and slightly bitter taste. The extraction process produced 25.55% yield. This result has met the standard of the Indonesian Herbal Pharmacopoeia (Depkes RI, 2008). The curcumin content was detected in the extract, as shown in Figure 1. While curcumin was found to be a major content in the extract, the other curcumin derivatives (i.e., demethoxycurcumin and bisdemethoxycurcumin) were not detected.

Curcumin is one of major chemical content in EEZC. Curcumin reported to have immunomodulatory activity (Varalakshmi et al., 2008). Previous study reported the significantly increase of IL-12 levels in curcumin-treated animals on day 10 and 20, after treatment. It is also found that curcumin induce generation of ROS which important in the immune respon (Varalakshmi et al., 2008). Curcuminoid (cassumunin A and cassumunin B) isolated from *Z. cassumunar* was found to have a protective effect on living cells suffering from oxidative stress (Nagano et al., 1997).

Beside curcumin, essential oil is also reported is one of main compound in *Zingiber cassumunar* rhizome. The highly essential oil content give specific odor of *Z. cassumunar* rhizome and extract. Several studies of phytochemical compounds and biological activity of *Z. cassumunar* Roxb had been reported the main component of *Z. cassumunar* rhizome essential oil were triquinacene 1,4-bis (methoxy), (Z)-ocimene and terpinen-4-ol (Bua-in and Paisooksantivatana, 2009). The previous studies on its rhizome also found several phenylbutenoid compounds, curcuminoid, and sesquiterpene (zerumbon) (Nakamura et al., 2009).

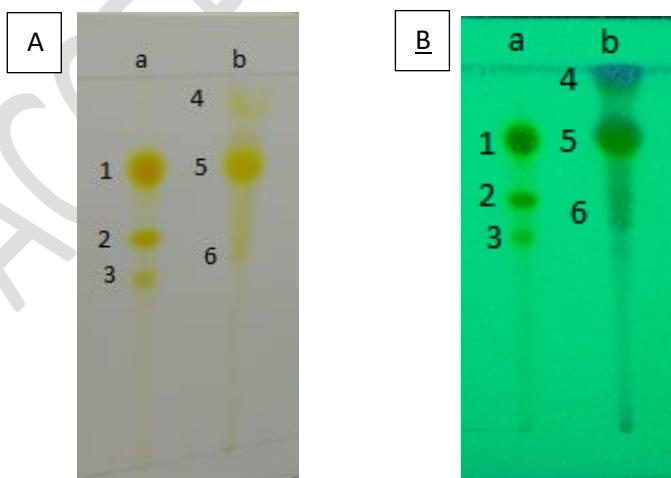
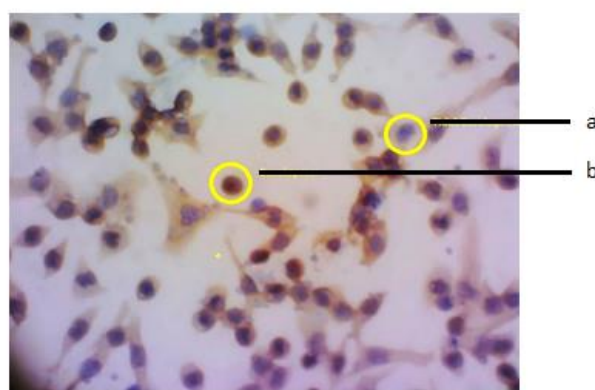


Figure 1. The TLC profile of curcuminoids standard (a) and *Zingiber cassumunar* ethanolic extract (b), detected in daylight (A) and UV 254 nm (B).

191 **Expression of IL-10**

192 Indirect immunocytochemistry was employed to detect the expression of interleukin in the
193 macrophage. The specific antibody of IL-10 interacted with interleukin-10 in the cells and attached
194 to the secondary antibody. During the detection process, the secondary antibody attached to DAB as
195 the chromogen, and the expression appeared as brown stains on the cytoplasm area. Meanwhile, the
196 cells with negative expression appeared in blue as the result of counterstaining. The
197 immunocytochemistry of the macrophage is shown in Figure 2. The percentage of the IL-10
198 expression is shown in Table 1.

199



200

201 Figure 2 The immunocytochemistry of IL-10 expression in macrophage cells after treatment with
202 ethanol extract of *Zingiber cassumunar* which observed with 400x magnification (a)
203 macrophage with no expression of interleukin (b), macrophage with positive expression of
204 interleukin
205

206 Table 1 The percentage of IL-10 expression on the macrophage cells of BALB/c mice treated with
207 ethanol extract of *Zingiber cassumunar*.

Groups	Mean \pm SD
Normal	45.65 \pm 1.92%*
Negative control	51.86 \pm 1.42%
Treatment Dose of 1.25mg/20gBW	55.91 \pm 3.07%
Treatment Dose of 2.5mg/20gBW	63.68 \pm 2.93%*
Treatment Dose of 5mg/20gBW	68.65 \pm 4.42%*

208 *significant difference with negative control (p<0.05)

209

210 The treatment of ethanol extract of *Zingiber cassumunar* increased the expression of IL-10,
211 affirming the potential of EEZC as an immunomodulator. IL-10 was expressed by macrophages and
212 other dendritic cells (DC) as a response to microbial infection. The increase of IL-10 expression may
213 be attributable to the activation of extracellular signal-regulated kinase 1 (ERK1) and ERK2 (Saraiva
214 and O'Garra, 2010). Such an increase will activate the specific response of the immune system and

215 inhibit the nonspecific response. IL-10 has been identified as an inhibitor of the synthesis of
 216 inflammatory mediators and pro-inflammatory cytokines that play a role in modulating fever and
 217 sickness (Harden et al., 2013). The present study also found that the expression of IL-10 in negative
 218 control group increased significantly compared to normal, which could be caused by the tween 80
 219 effect. The previous study also reported that polysorbate (tween) 80 could increase the immune
 220 response (Maggio, 2012).

221 The increased expression of IL-10 was proportional to the administered dose. The higher the
 222 treatment dose, the higher the expression of IL-10 was. However, an extremely high level of IL-10
 223 can inhibit chemokine production and prevent its role in directing lymphocytes to the lymph nodes,
 224 as in mycobacterial infection, resulting in a failure to recruit and induce Th1 cell differentiation
 225 (Couper et al., 2008). Therefore, IL-10 has both immunosuppressive and immunostimulatory
 226 properties (Acuner-ozbabacan et al., 2014).

227 The regulation of IL-10 expression involves the enhancement or silencing of IL10 transcription
 228 and is regulated by certain transcription factors activated by discrete signal-transduction pathways.
 229 Following transcription, the post-transcriptional mechanisms exist and involves many of the
 230 molecular events leading to IL-10 expression (Saraiva and O'Garra, 2010). Some molecule of
 231 *Zingiber cassumunar* could be involves and affect the transcriptional process of IL-10 and leading on
 232 increasing level of IL-10.

233

234 **Expression of Interleukin-12**

235 Interleukin-12 (IL-12) is a pro-inflammatory cytokine that induces the production of interferon-
 236 γ (IFN- γ), and leading to the differentiation of T helper 1 (TH1) cells and connected the link of innate
 237 immunity and adaptive immunity. Dendritic cells (DCs) and macrophages produce IL-12 in response
 238 to pathogens and infection (Trinchieri, 2003). Production of IL-12 is strongly regulated by positive
 239 and negative regulatory mechanisms. Microorganism products including bacteria, intracellular
 240 parasites, fungi, double-stranded RNA, bacterial DNA and oligonucleotides are strong inducers of
 241 IL-12 production by macrophages, monocytes, neutrophils and DCs. The LPS was used in this study
 242 to activate the macrophage to produce the IL-12. The quantitative analysis of the expression of IL-
 243 12 after the administration of EEZC is shown in Table 2.

244

245 Table 2 The expression of IL-12 in macrophage cells of BALB/c mice treated with ethanol extract of
 246 *Zingiber cassumunar*

Groups	Mean \pm SD
Normal	64.63% \pm 9.763
Negative Control	66.39% \pm 1.603
Treatment Dose of 1.25mg/20g BW	51.56% \pm 4.528*
Treatment Dose of 2.5mg/20g BW	70.62% \pm 3.469

247 *) significant difference with negative control (p<0.05)

248

249 The study found that the IL-12 expression in the negative control group was not significantly
250 different from the normal group, indicating that the solvent (tween 80) does not affect the immune
251 response. Tween 80 could stimulate the immunogenicity (Maggio, 2012) as shown by the increasing
252 of IL-10 in the present study. But the dose used in this study is not enough to increase the IL-12
253 expression. The treatment of EEZC at a dose of 1.25mg/20g BW induced lower IL-12 expression
254 than the negative control. In other words, the lower the dose, the less effective the active compound
255 of EEZC in increasing the IL-12 expression. When the dose increased, the IL-12 expression was also
256 found to be elevated.

257 The immunomodulatory effects of EEZC might be caused by the presence of curcumin as an
258 active compound. Curcumin has been reported to increase the immune response of the cells (Nagano
259 et al., 1997; Nurkhasanah et al., 2017). The previous study also found that curcumin treatment
260 elevated the IL-12 level in mice (Varalakshmi et al., 2008). The increasing level of IL-12 in the
261 treatment could be caused by the capacity of curcumin in increasing of ROS and NO (Nurmasari et
262 al., 2014; Rahmawati, 2013). ROS is known to regulate the IL-12 generation. Another studies have
263 revealed that curcumin stimulates T cell, B cell, neutrophil, NK cell, and dendritic cell (Nurmasari et
264 al., 2014).

265 The essential oil, which emitted a special odor, was also identified in EEZC (Bhuiyan et al.,
266 2008). The presence of essential oils in EEZC has also been reported to boost the immune response,
267 including the phagocytic activity of macrophages (Chairul et al., 2009; Nakamura et al., 2009). The
268 active compound from volatile oil which successfully identified as immunomodulatory compound
269 are phenilbutenoids compound (Chairul et al., 2009).

270 The treatment of EEZC increased IL-12 expression, activating T cells and stimulating the
271 production of IFN- γ , which lead to macrophage activation and the secretion of reactive oxygen
272 species (ROS) that eliminate infections (Abbas et al., 2017). Furthermore, this treatment can intensify
273 the phagocytic activity of macrophage (Nurkhasanah et al., 2017).

274

275 **Expression of Interleukin-14**

276 IL-14 was first known as a high-molecular-weight B-cell growth factor and originally
277 identified as a B cell growth factor (Shen et al., 2006). It is produced by T cells and B-cells. IL-14
278 binds and signals through a 90-kDa receptor expressed on activated B cells to promote B-cell
279 proliferation (Akdis et al., 2016). High level of IL-14 can enhance B-cell proliferation and expand a

280 subpopulation of memory B cells (Leca et al., 2008), and if followed by the secretion of antibody, it
281 can eliminate the invader. The expression of IL-14 in EEZC-treated mice is shown in Table 3.

282

283 Table 3 The expressions of Interleukin-14 in mice treated with ethanol extract of *Zingiber*
284 *cassumunar*

Groups	Expressions (X ± SD)
Normal	59.19 ± 3.07%
Negative control	61.24 ± 1.51%
Treatment Dose of 1.25 mg/20g BW	57.02 ± 1.94%*
Treatment Dose of 2.5 mg/20g BW	67.41 ± 6.60%
Treatment Dose of 5 mg/20g BW	71.07 ± 1.30%*

285 (*) showed significant difference with negative control (p<0.05)

286

287 The previous research proposes increasing the expression of IL-14 by the treatment of some
288 herbal medicine extract (Nurkhasanah, 2015). The treatment of anthocyanin-rich rosella extract
289 increases the IL-10 and IL-14 expressions in vitro. The present research also found that the treatment
290 of EEZC increased the IL-10, IL-12, and IL-14 expressions after LPS induction. This induction
291 stimulated the immune response because LPS was recognized as endotoxin consisting of a lipid and
292 a polysaccharide found on the outer membrane of gram-negative bacteria. The activity of EEZC in
293 increasing of IL-10, IL-12 and IL-14 suggested the potency of this extract in inducing the immune
294 system in both innate and adaptive immunity.

295 A previous study on Zingiberaceae family, including *Curcuma mangga*, *Kaempferia*
296 *angustifolia*, and *Zingiber cassumunar*, highlights that *Zingiber cassumunar* has the highest
297 immunomodulatory activity (Chairul et al., 2009), which may be caused by the active compound of
298 curcumin and essential oil. The major compound found in *Z. cassumunar* essential oil was phenyl
299 butanoic compound. Furthermore, a toxicity study states that *Zingiber cassumunar* extract has no
300 observable adverse effect and it is well-tolerated for both acute and chronic studies (Koontongkaew
301 et al., 2014).

302

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CONCLUSION

304 The ethanol extract of *Zingiber cassumunar* has immunomodulatory activity through increasing
305 level of IL-10, IL-12 and IL-14 cytokines and suggested the potency of extract to induce both innate
306 and adaptive immunity.

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